



SUPPLEMENTARY MATERIAL TO
**Chromium(VI) removal from aqueous solutions using a
polyethylenimine–epichlorohydrin resin**

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THE RESIN

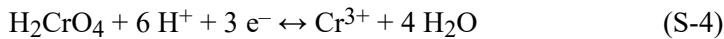
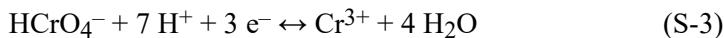
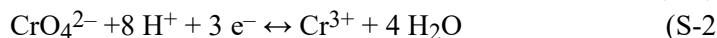
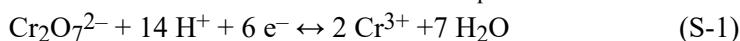
The non-crystalline spherical particles of the resin had an average diameter of 80 µm and specific surface area of 0.410 m² g⁻¹. The resin was found to be stable up to ca. 180 °C.

The XPS examination of the resin showed the existence of both ternary (nitrogen binding energy (BE) 399.2 eV) and quaternary (BE: 401.4 eV) amino-groups in the relation 45 to 55 %.^{1–3} These groups can justify the anion-sorption properties of the material. The quaternary amine groups can exchange anions over the whole pH range, whereas the ternary ones show, because of their protonation, a higher metal uptake capacity at low pH values. A small number of COO⁻-groups (ca. 8 %) were also observed by XPS. Additional information about the characterization of the resin is given in a previous publication.⁴

Influence of the resin on the chromates solution pH

The equilibrium pH (pH_{equil}) of the investigated Cr-solutions as a function of the equilibrium concentration (*c*_{equil}) is shown in Fig. S-1.

The following reactions can lead to an increase of pH_{equil}:⁵



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During Cr uptake, a change in the color of the resins (from yellowish to black) at all pH values was observed.

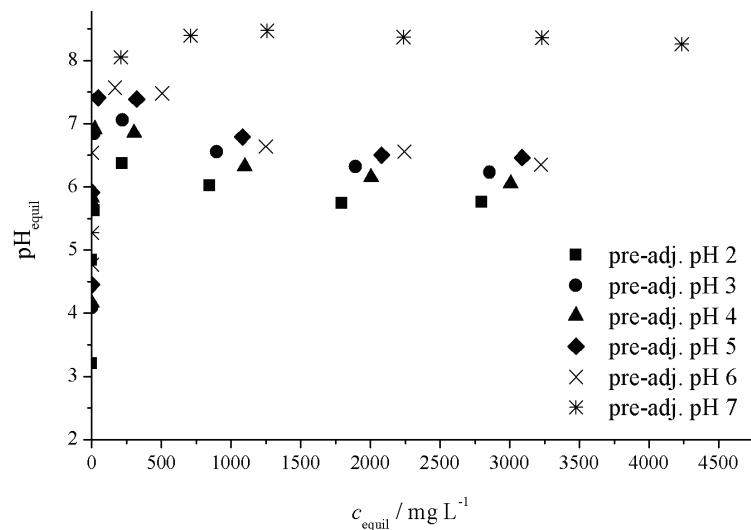


Fig. S-1. Equilibrium pH of Cr(VI) solution *vs.* c_{equl} .

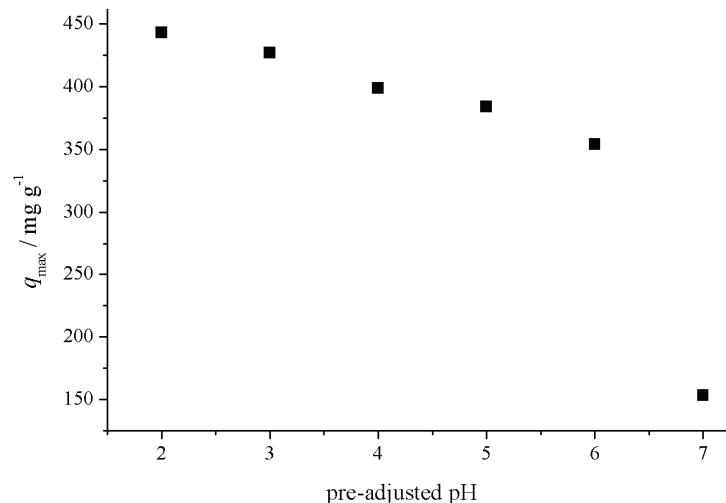


Fig. S-2. Experimental maximum sorption capacity, q_{max} , from solutions of different pre-adjusted pH values.

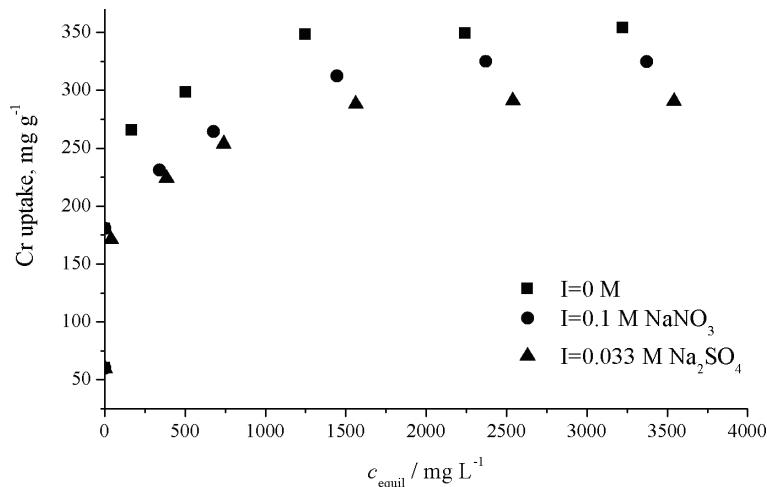


Fig. S-3. Isotherms for Cr(VI)-removal by the resin from aqueous solutions of pre-adjusted pH 6 in the presence of background electrolytes.

TABLE S-I. Adsorption isotherm models (q_{equil} and c_{equil} : equilibrium concentration of Cr(VI) in solid and liquid phase, respectively; q_{max} : the maximum sorption capacity in mg g^{-1} ; K : the equilibrium constant; n and m : parameters characterizing the system heterogeneity)

Isotherm model	Equation
Langmuir (L)	$q_{\text{equil}} = q_{\text{max}}Kc_{\text{equil}}/(1 + Kc_{\text{equil}})$
Redlich-Peterson (RP)	$q_{\text{equil}} = q_{\text{max}}Kc_{\text{equil}}/(1 + (Kc_{\text{equil}})^n)$
Langmuir-Freundlich (LF)	$q_{\text{equil}} = q_{\text{max}}(Kc_{\text{equil}})^n/(1 + (Kc_{\text{equil}})^n)$
Toth (T)	$q_{\text{equil}} = q_{\text{max}}Kc_{\text{equil}}/[1 + (Kc_{\text{equil}})^n]^{1/n}$

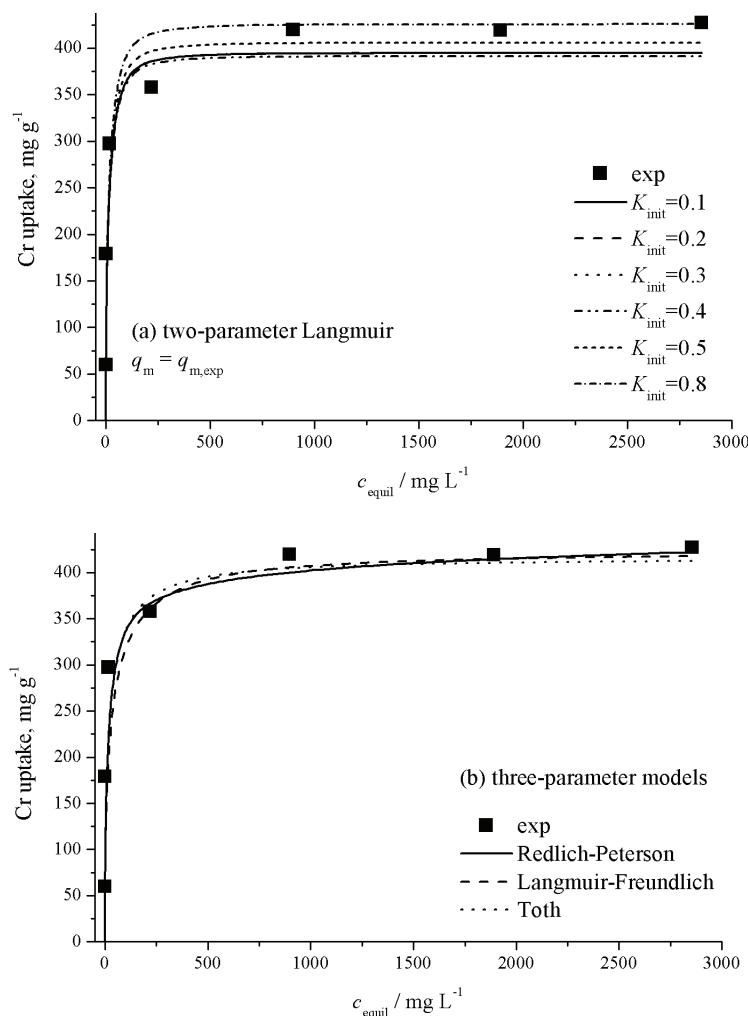


Fig. S-4. Experimental (symbols) and calculated (lines) isotherms for Cr(VI) sorption from aqueous solutions pre-adjusted pH 3 by the resin: a) the sensitivity of the Langmuir model to an initial guess of the estimated parameter K ($q_{\text{init},\text{max}}$ was set equal to $q_{e,\text{max}} = 427.05 \text{ mg g}^{-1}$); b) three-parameter models.

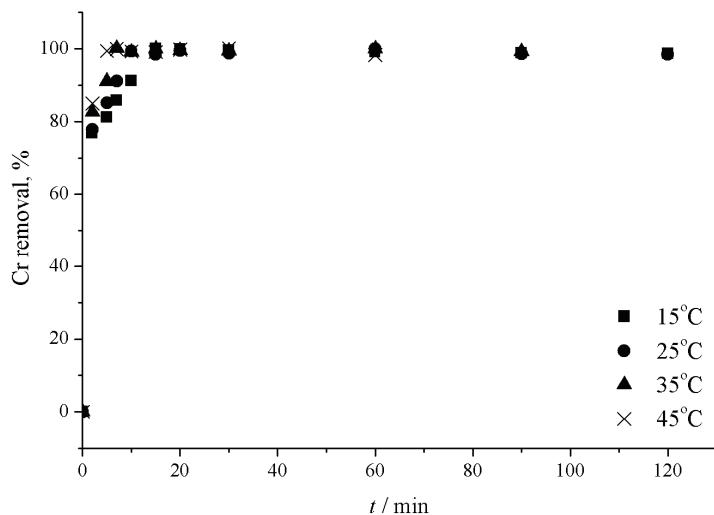


Fig. S-5. Time dependence of chromium removal onto the resin at different temperatures.

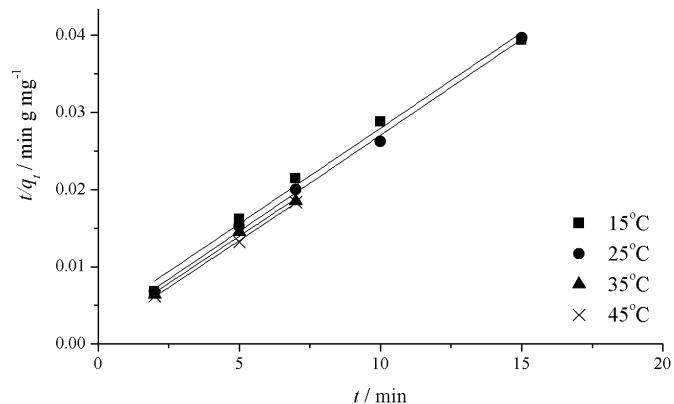


Fig. S-6. Pseudo-second order sorption kinetics of Cr(VI) onto the resin at various initial temperatures.

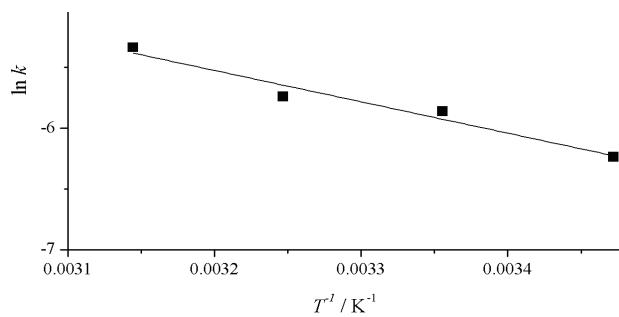


Fig. S-7. Plot of $\ln k$ against temperature for Cr(VI) sorption onto the resin.

TABLE IV. A comparison of sorption capacity for the Cr(VI)-sorption by selected sorbents reported in the literature

Sorbent	pH	Sorption capacity, mg g ⁻¹	Ref.
<i>Pterocladia capillacea</i>	1	12	6
Surfactant-modified montmorillonite	3	18	7
Resin Lewatit MP 64	5	20.8	8
Resin Lewatit MP 500	6	21.3	8
Amberlite IRA96 ion-exchange resin	3	23.9	9
Dowex 1×8 ion-exchange resin	3	28.1	9
Untreated <i>Coriolus versicolor</i>	2	44.2	10
Ethylenediamine-modified cross-linked magnetic chitosan resin	2	51.8	11
Chemically modified <i>Sargassum sp.</i>	2	58.4	12
Ethylenediamine (EDA)-functionalized magnetic polymers	2.5	61.4	13
Aakaganeite (b-FeO(OH))	5.5	80	14
Chitin	2	101.2	15
<i>Rhizopus arrhizus</i>	2	108.9	16
Anion-exchange resin D314	4.5	120.5	17
Chitosan coated with poly-3-methyl thiophene	2	127.6	18
Stabilized iron nanoparticles	1	131.6	19
	3	123.4	
	5	96.2	
	7	112.4	
<i>Staphylococcus xylosus</i>	1	143	20
Anion-exchange resin D301	4.5	152.5	21
Anion-exchange resin D354	4.5	156.2	22
Tannery residual biomass	2	217	23
Fe-crosslinked chitosan complex	4.7	295	24
Polyethylenimine modified aerobic granules sludge	5.2	348.1	25
Aerobic granules functionalized with polyethylenimine	5.5	401.5	26
Polyethylenimine–epichlorohydrin resin	2	443	This work
Chitosan grafted with acrylic acid	4	518	27
Chitosan	4	655	27
Chitosan grafted with acrylamide	4	935	27

REFERENCES

1. S. Kundu, W. Xia, W. Busser, M. Becker, D. A. Schmidt, M. Havenith, M. Muhler, *Phys. Chem. Chem. Phys.* **12** (2010) 4351
2. J. R. Pels, F. Kapteijn, J. A. Moulijn, Q. Zhu, K. M. Thomas, *Carbon* **33** (1995) 1641
3. E. Baumgarten, A. Fiebes, A. Stumpe, F. Ronkel, J. W. Schultze, *J. Mol. Catal. A: Chem.* **113** (1996) 469
4. S. Sarri, P. Misaelides, D. Zamboulis, X. Gaona, M. Altmaier, H. Geckeis, *J. Radioanal. Nucl. Chem.* **307** (2016) 681

5. J. C. Bailar Jr., H. J. Emeléus, R. Nyholm, A. F. Trotman-Dickenson, *Comprehensive Inorganic Chemistry*, Vol. 3, Pergamon Press, Oxford, 1973, p. 637
6. A. El Nemr, A. El-Sikaily, A. Khaled, O. Abdelwahab, *Arabian J. Chem.* **8** (2015) 105
7. M. C. Brum, J. L. Capitaneo, J. F. Oliveira, *Miner. Eng.* **23** (2010) 270
8. E. Pehlivan, S. Cetin, *J. Hazard. Mater.* **163** (2009) 448
9. S. Edebali, E. Pehlivan, *Chem. Eng. J.* **161** (2010) 161
10. R. Sanghi, N. Sankararamakrishnan, B. C. Dave, *J. Hazard. Mater.* **169** (2009) 1074
11. X. Hu, J. Wang, Y. Liu, X. Li, G. Zeng, Z. Bao, X. Zeng, A. Chen, F. Long, *J. Hazard. Mater.* **185** (2011) 306
12. L. Yang, J. P. Chen, *Bioresour. Technol.* **99** (2008) 297
13. Y. G. Zhao, H. Y. Shen, S. D. Pan, M. Q. Hu, *J. Hazard. Mater.* **182** (2010) 295
14. N. K. Lazaridis, D. N. Bakoyannakis, E. A. Deliyanni, *Chemosphere* **58** (2005) 65
15. Y. Sağ, Y. Aktay, *Process Biochem.* **36** (2000) 157
16. Z. Aksu, E. Balibek, *J. Hazard. Mater.* **145** (2007) 210
17. M. Ghiaci, R. Kia, A. Abbaspur, F. Seyedeyn-Azad, *Sep. Purif. Technol.* **40** (2004) 285
18. S. Hena, *J. Hazard. Mater.* **181** (2010) 474
19. A. Bampaiti, F. Noli, P. Misaelides, *J. Radioanal. Nucl. Chem.* **298** (2013) 909
20. M. Ziagova, G. Dimitriadis, D. Aslanidou, X. Papaioannou, E. Litopoulou Tzannetaki, M. Liakopoulou-Kyriakides, *Bioresour. Technol.* **98** (2007) 2859
21. N. Zhao, N. Wei, J. Li, Z. Qiao, J. Cui, F. He, *Chem. Eng. J.* **115** (2005) 133
22. T. Shi, Z. Wang, Y. Liu, S. Jia, D. Changming, *J. Hazard. Mater.* **161** (2009) 900
23. J. Anandkumar, B. Mandal, *J. Hazard. Mater.* **186** (2011) 1088
24. A. C. Zimmermann, A. Mecabô, T. Fagundes, C. A. Rodrigues, *J. Hazard. Mater.* **179** (2010) 192
25. X. F. Sun, C. Liu, Y. Ma, S. G. Wang, B. Y. Gao, X. M. Li, *Colloids Surf. B* **82** (2011) 456
26. X. F. Sun, Y. Ma, X. W. Liu, S. G. Wang, B. Y. Gao, X. M. Li, *Water Res.* **44** (2010) 2517
27. G. Z. Kyzas, M. Kostoglou, N. K. Lazaridis, *Chem. Eng. J.* **152** (2009) 440.