



SUPPLEMENTARY MATERIAL TO
**Antimicrobial and anti-tubercular activities of isolates and
semi-synthetic derivatives of lichen *Ramalina leiodea* (Nyl.) Nyl.**

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EXPERIMENTAL

Antimicrobial activities

The determinations of both the antibacterial and antifungal activities for measuring the antimicrobial properties were realized according to the standard well plate method.¹ For the antibacterial activities, all the selected test strains (*Salmonella enterica* ATCC 35664, *Pseudomonas aeruginosa* ATCC 15442, *Bacillus subtilis* ATCC 23857 and *Staphylococcus aureus* ATCC 25923) were initially activated and grown in nutrient agar. Whereas, for anti-fungal studies, the test strain used, *Candida albicans*, was grown on potato dextrose agar medium. All compounds were re-dissolved in DMSO to obtain a final concentration of 1 mg mL⁻¹, which were used as stock solutions. The compounds were used for activity studies and the concentration of each sample was 1 µg mL⁻¹ along with standard and control. The media, Petri dishes were autoclaved at 121 °C for 15 min. After sterilization, the agar plates were prepared by pouring 25 mL of agar medium followed by incubation at room temperature for 30 min for solidification under a sterile environment. These plates were inoculated with 60 µL of test inoculums using sterile cotton swabs. Wells 8 mm in diameter were made with a sterile cork borer and into each well exactly 100 µL of sample were loaded. The control and standard were placed in separate wells. The plates were initially incubated for 20–30 min at 4 °C to allow the compounds to diffuse into the agar, and then subsequently incubated for 24 h at 37 °C for the bacteria and 48 h at 28 °C for the fungi. The zone diameters were measured in mm using a calibrated scale. The experiments were conducted in triplicate with aliquots to minimize the deviations and the average values are reported.

The compounds having better anti-microbial activities were selected for the minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) studies against *Salmonella enterica*, *P. aeruginosa*, *B. subtilis*, *S. aureus* and *C. albicans* according to a reported method.² The concentrations of test samples were serially diluted from 1 to 0.001 µg mL⁻¹ and one tube without drug served as the control. All the tubes were inoculated with 1 mL of respective cultures having an OD of 0.2 (≈McFarland standard) and the tubes were incubated at 37 °C for 12–16 h. The turbidity of each tube was measured with respect to the control tube. The MIC values are defined as the lowest concentration of a compound at which

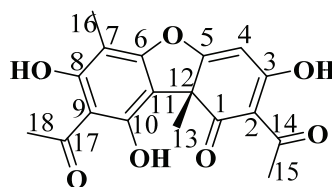
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growth was completely inhibited. After incubation, the culture from each tube was plated in nutrient agar to evaluate the *MBC* value. The concentration at which the cells were all dead was defined as the *MBC*.

Anti-tubercular activity

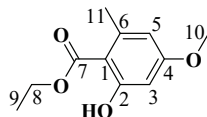
The *in vitro* anti-tubercular activity assessment of all compounds was tested against the *Mycobacterium tuberculosis* H37Rv strain ATCC 25177 using the microplate alamar blue assay (MABA)^{2,3} in three sets ($n = 3$). The minimum inhibitory concentration (*MIC*) was determined by the color change from blue to pink, which indicate no bacterial growth and growth, respectively.

ANALYTICAL AND SPECTRAL DATA FOR THE COMPOUNDS



1

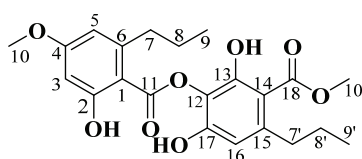
Usnic acid (1). Yield: 236 mg (0.16 %, based on total weight of the lichen material); m.p.: 201–202 °C, Anal. Calcd for C₁₈H₁₆O₇: C, 63.15; H, 6.26 %. Found C, 63.16; H, 6.26 %; IR (KBr, cm⁻¹): 2923, 2858, 1687, 1626, 1543, 1462, 1368, 1293, 1192, 1138, 1071, 1037, 966, 843; ¹H-NMR (400 MHz, CDCl₃, δ / ppm): 1.76 (3H, s, Me-13), 2.11 (3H, s, Me-16), 2.67 (3H, s, Me-15), 2.68 (3H, s, Me-18), 5.98 (1H, s, H-4), 11.03 (1H, s, OH-10), 13.31 (1H, s, OH-8); ¹³C-NMR (100 MHz, CDCl₃, δ / ppm): 7.6 (C-16), 27.9 (C-13), 31.3 (C-18), 32.2 (C-15), 59.1 (C-12), 98.4 (C-4), 101.6 (C-7), 104.0 (C-11), 105.3 (C-2), 109.3 (C-9), 155.2 (C-6), 157.5 (C-10), 163.9 (C-8), 179.4 (C-5), 191.7 (C-3), 198.1 (C-1), 200.4 (C-14), 201.8 (C-17); MS (EI, 70 eV, m/z (%): 343 [M–H⁺] (100), 345 [M+H⁺] (100); HRMS-FAB (m/z [M+H⁺]) Calcd. for C₁₈H₁₆O₇: 344.09. Found: 344; UV/Vis (EtOH, λ_{\max} / nm: 220. *R_f* (hexane–DCM, 9:1): 0.4.



2

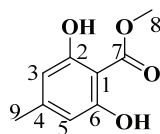
Ethyl everninate (2). Yield: 153 mg (0.10 %, based on total weight of the lichen material); m.p.: 74–76 °C, Anal. Calcd for C₁₁H₁₄O₄: C, 62.85; H, 6.71 %. Found: C, 62.87; H, 6.82 %; IR (KBr, cm⁻¹): 2920, 2855, 1625, 1462, 1371, 1280, 1243, 1164, 757; ¹H-NMR (400 MHz, DMSO-*d*₆, δ / ppm): 0.95 (3H, t,

$J = 8$ Hz, Me-9), 2.22 (3H, *s*, Me-11), 3.02 (H, *s*, OH-2), 3.83 (3H, *s*, OMe-10), 4.37 (2H, *m*, OCH₂), 6.33 (1H, *d*, $J = 1.2$ Hz, H-5), 6.37 (1H, *d*, $J = 1.2$ Hz, H-3); ¹³C-NMR (100 MHz, acetone-*d*₆, δ / ppm): 14.47 (C-9), 25.85 (C-11), 54.96 (C-10), 55.8 (C-8), 99.7 (C-1), 111.1 (C-3/5), 149.1 (C-6), 165.2 (C-2), 167.2 (C-4), 174.0 (C-7); MS (EI, 70 eV, m/z (%)): 209.1 [M-H⁺] (100), 211 [M+H⁺] (100); HRMS-FAB: (m/z [M + H⁺]) Calcd. for C₁₁H₁₄O₄: 210.09. Found: 210; UV/Vis (EtOH, λ_{\max} / nm: 219; R_f (hexane-DCM, 7:3): 0.4.



3

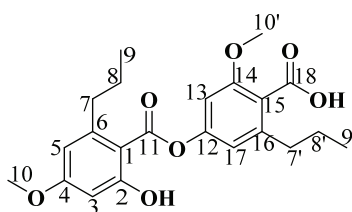
Scrobiculin (3). Yield: 911 mg (0.61 %, based on total weight of the lichen material), m.p.: 134–136 °C, Anal. Calcd for C₂₂H₂₆O₈: C, 63.15; H, 6.26. Found C, 63.16; H, 6.26. IR (KBr, cm⁻¹): 2958, 2926, 2864, 1657, 1619, 1581, 1503, 1455, 1420, 1342, 1239, 1160, 1129, 1088, 1037, 830, 757; ¹H-NMR (400 MHz, DMSO-*d*₆, δ / ppm): 0.88–0.97 (6H, *m*, Me-9, 9'), 1.59–1.65 (4H, *dd*, CH₂-8, 8'), 2.51 (1H, *t*, OH-2), 2.82–2.87 (4H, *dd*, $J_1 = 4$ Hz, $J_2 = 4$ Hz CH₂-7, 7'), 3.78 (3H, *s*, OMe-10), 3.84 (3H, *s*, OMe-10'), 6.39–6.41 (2H, *dd*, $J_1 = 0.8$ Hz, $J_2 = 1$ Hz H-3, 5), 6.61 (1H, *s*, H-16), 10.50 (1H, *s*, OH-17), 11.85 (1H, *s*, OH-13); ¹³C-NMR (100 MHz, DMSO-*d*₆, δ / ppm): 14.5 (C-9), 14.6 (C-9'), 24.8 (C-8), 25.2 (C-8'), 36.9 (C-7), 37.8 (C-7'), 55.8 (C-10), 56.5 (C-10'), 99.5 (C-3), 106.2 (C-1), 108.6 (C-16), 108.8 (C-14), 109.5 (C-5), 125.2 (C-12), 144.1 (C-15), 146.0 (C-6), 154.3 (C-13), 154.7 (C-17), 160.6 (C-2), 162.9 (C-4), 166.7 (C-11), 172.7 (C-18); MS (EI, 70 eV, m/z (%)): 417 [M-H⁺] (100), 418.9 [M+H⁺] (100); HRMS-FAB (m/z [M + H⁺]) Calcd. for C₂₂H₂₆O₈: 418.44. Found: 418, UV/Vis (EtOH, λ_{\max} / nm: 217.5; R_f (DCM-EA, 7:3): 0.6.



4

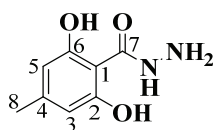
Methyl 2,6-dihydroxy-4-methylbenzoate (4). Yield: 4.05 g (2.70 %, based on total weight of the lichen material); m.p.: 143–144 °C, Anal. Calcd for C₉H₁₀O₄: C, 59.34; H, 5.53 %. Found C, 59.36, H, 5.52 %; IR (KBr, cm⁻¹): 3866, 3746, 3370, 3312, 2982, 2955, 2854, 1894, 1642, 1581, 1503, 1447, 1383, 1318, 1265, 1197, 1162, 1114, 1059, 1029, 996, 950, 835, 800, 753, 700, 620, 575; ¹H-NMR (400 MHz, DMSO-*d*₆, δ / ppm): 2.23 (3H, *s*, Me-9), 3.75 (3H, *s*, OMe-8), 6.12

(2H, *d*, *J* = 2.0 Hz, H-3, 5), 9.94 (1H, *s*, OH-6), 10.65 (1H, *s*, OH-2); ¹³C-NMR (100 MHz, DMSO-*d*₆, δ / ppm): 22.5 (C-9), 52.2 (C-8), 100.8 (C-1), 107.9 (C-5), 110.6 (C-3), 141.2 (C-4), 161.5 (C-2/6), 170.6 (C-7). MS (EI, 70 eV, *m/z* (%)): 183.0 [M + H⁺] (100); HRMS-FAB (*m/z* [M+H⁺]) Calcd. for C₉H₁₀O₄: 182.18. Found: 182; UV/Vis (EtOH, λ_{\max} / nm): 219.5; *R_f*(hexane–EA, 1:1): 0.6.



5

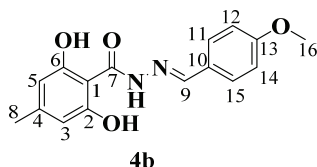
4-[(2-hydroxy-4-methoxy-6-propylbenzoyl)oxy]-2-methoxy-6-propylbenzoic acid (5). Yield: 11 mg (0.007 %, based on total weight of the lichen material); m.p.: 142–143 °C; Anal. Calcd for C₂₂H₂₆O₇: C, 65.66; H, 6.51 %. Found: C, 65.66; H, 6.62 %; ¹H-NMR (400 MHz, CDCl₃, δ / ppm): 0.93–1.05 (6H, *m*, Me-9, 9'), 1.66–1.80 (4H, *m*, CH₂-8, 8'), 2.95–3.04 (4H, *m*, CH₂-7, 7'), 3.85 (3H, *s*, OMe-10), 3.91 (3H, *s*, OMe-10'), 6.40 (2H, *s*, H-13, 17), 6.46 (2H, *s*, H-3, 5), 11.18 (1H, *s*, OH-18), 11.73 (1H, *s*, OH-2); ¹³C-NMR (100 MHz, CDCl₃, δ / ppm): 14.3 (C-9/9'), 24.81 (C-8'), 25.18 (C-8), 38.8 (C-7'), 39.1 (C-7), 55.4 (C-10), 56.0 (C-10'), 99.8 (C-3), 104.5 (C-1), 106.2 (C-15), 107.1 (C-13), 110.9 (C-5), 146.8 (C-17), 148.7 (C-16), 151.6 (C-6), 156.1 (C-12), 156.9 (C-14), 164.4 (C-4), 165.5 (C-2), 168.9 (C-11), 173.7 (C-18); MS (EI, 70 eV, *m/z* (%)): 403.53 [M–H⁺] (100); HRMS-FAB: (*m/z* [M + H⁺]) Calcd. for C₂₂H₂₆O₇: 402.17. Found: 402.53; UV/Vis (EtOH, λ_{\max} / nm): 212.5; *R_f*(hexane–EA, 1:1): 0.4.



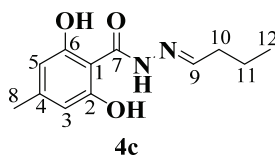
4a

2,6-Dihydroxy-4-methylbenzohydrazide (4a). Yield: 77 %; m.p.: 153–154 °C; Anal. Calcd for C₈H₁₀N₂O₃: C, 52.74; H, 5.53; N, 15.38 %. Found C, 52.64; H, 5.52; N, 15.48 %; IR (KBr, cm⁻¹): 2917, 2564, 1621, 1498, 1458, 1367, 1325, 1266, 1216, 1173, 1001, 919, 841, 800, 735, 688; ¹H-NMR (400 MHz, CDCl₃, δ / ppm): 2.57 (3H, *s*, Me), 5.07 (2H, *s*, NH₂), 6.42 (2H, *s*, Ar-H), 7.41 (1H, *s*, NH), 10.05 (2H, *s*, Ar-OH); ¹³C-NMR (100 MHz, CDCl₃, δ / ppm): 24.4 (C-8), 99.7 (C-1), 112.2 (C-3/5), 144.7 (C-4), 164.0 (C-2/6), 166.5 (C-7); MS (EI, 70

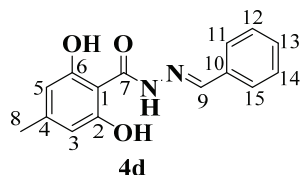
eV,; m/z (%): 183.35 [M + H⁺]; HRMS-FAB (m/z [M + H⁺]) Calcd. for C₈H₁₀N₂O₃: 182.18. Found: 182.35.



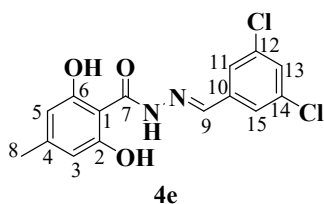
(*E*)-2,6-Dihydroxy-*N'*-(4-methoxybenzylidene)-4-methylbenzohydrazide (**4b**). Yield: 68 %, m.p.: 120–121 °C. Anal. Calcd for C₁₆H₁₆N₂O₄: C, 63.99; H, 5.37, N, 9.33 %. Found: C, 64.00; H, 5.32; N, 9.37 %; IR (KBr cm⁻¹): 3205, 3054, 2843, 1655, 1804, 1509, 1462, 1428, 1379, 1340, 1300, 1250, 1171, 1096, 1028, 960, 761, 668; ¹H-NMR (400 MHz, CDCl₃, δ / ppm): 2.38 (3H, *s*, Me), 3.84 (3H, *s*, OMe), 6.37–6.38 (2H, *d*, *J* = 4 Hz, Ar-H), 6.91–6.93 (2H, *m*, Ar-H), 7.60–7.62 (2H, *m*, Ar-H), 7.78 (1H, *s*, =CH), 9.89 (2H, *s*, Ar-OH); ¹³C-NMR (100 MHz, CDCl₃, δ / ppm): 20.4 (C-8), 55.4 (C-16), 111.5 (C-1), 114.2 (C-3/5), 126.6 (C-12/14), 128.7 (C-10), 138.3 (C-11/15), 143.8 (C-4), 147.3 (C-9), 161.2 (C-2/6), 164.4 (C-13), 174.1 (C-7); MS (EI, 70 eV, m/z (%)): 301.15 [M + H⁺] (100); HRMS-FAB (m/z [M + H⁺]) calcd. for C₁₆H₁₆N₂O₄: 300.31. Found: 300.15.



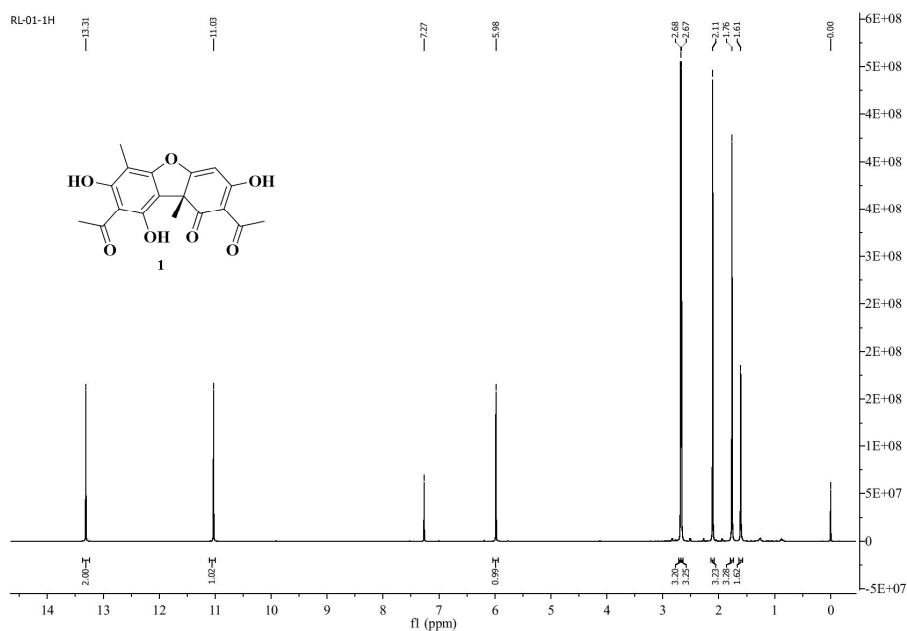
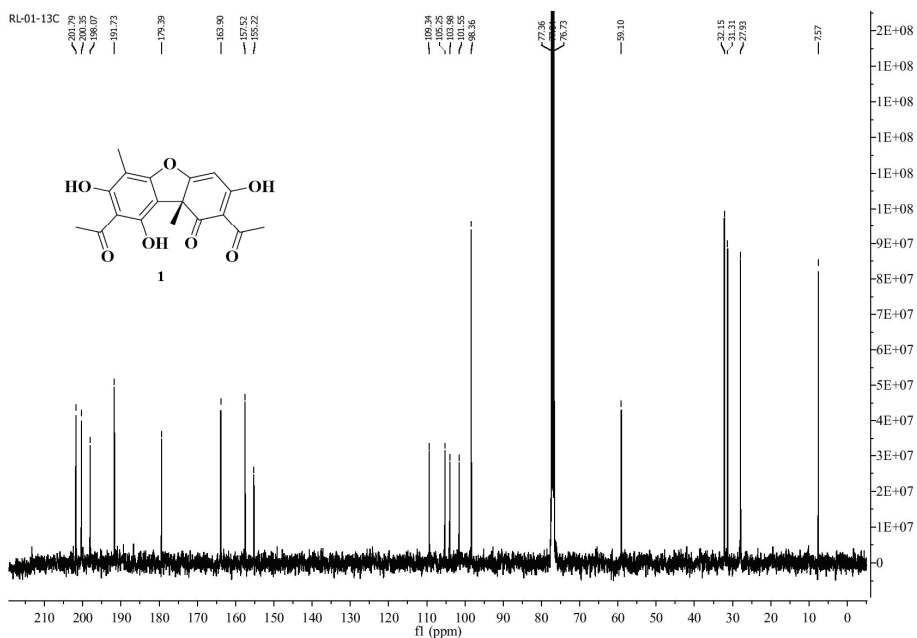
(*E*)-*N'*-Butylidene-2,6-dihydroxy-4-methylbenzohydrazide (**4c**). Yield: 70 %, m.p.: 110–111 °C; Anal. Calcd for C₁₂H₁₆N₂O₃: C, 61.00; H, 6.83, N, 11.89 %. Found: C, 61.00; H, 6.80; N, 11.89 %; IR (KBr, cm⁻¹): 3814, 3217, 3066, 2973, 2934, 1457, 1379, 1341, 1287, 1167, 1101, 846, 769; ¹H-NMR (400 MHz, CDCl₃, δ / ppm): 0.95–1.03 (3H, *m*, Me), 1.91–1.93 (2H, *m*, CH₂), 2.15 (3H, *s*, Me), 2.18–2.20 (2H, *m*, CH₂), 6.51 (*s*, 2H, Ar-H), 7.19–7.21 (*t*, *J* = 4 Hz, 1H, =CH), 7.83 (*s*, 1H, NH) 10.14 (*s*, 2H, Ar-OH); ¹³C-NMR (100 MHz, CDCl₃, δ / ppm): 14.2 (C-12), 19.8 (C-11), 22.7 (C-8), 29.8 (C-10), 106.5 (C-1), 113.9 (C-3/5), 130.5 (C-4), 149.5 (C-9), 155.8 (C-2/6), 168.6 (C-7); MS (EI, 70 eV, m/z (%)): 237.15 [M + H⁺]; HRMS-FAB (m/z [M + H⁺]) Calcd. for C₁₂H₁₆N₂O₃: 236.27. Found: 236.15.



(E)-N'-(Benzylidene)-2,6-dihydroxy-4-methylbenzohydrazide (**4d**). Yield: 64 %, m.p.: 98–99 °C. Anal. Calcd for C₁₅H₁₄N₂O₃: C, 66.66; H, 5.20, N, 10.36 %. Found: C, 66.68; H, 5.20, N, 10.40 %; IR (KBr, cm⁻¹): 3183, 3077, 2969, 2858, 1669, 1605, 1500, 1393, 1338, 1224, 1133, 1016, 948, 899, 758, 685; ¹H-NMR (400 MHz, CDCl₃, δ / ppm): 2.40 (3H, s, Me), 7.39–7.40 (3H, m, Ar-H), 7.67–7.68 (2H, m, Ar-H), 7.84 (1H, d, J = 4 Hz, =CH), 8.84 (1H, s, NH), 10.02 (2H, s, Ar-OH); ¹³C-NMR (100 MHz, CDCl₃, δ / ppm): 20.4 (C-8), 105.6 (C-1), 112.3 (C-3/5), 127.2 (C-12/14), 128.8 (C-11/13/15), 130.1 (C-10), 133.9 (C-4), 143.8 (C-9), 165.5 (C-2/6), 174.2 (C-7); MS (EI, 70 eV, m/z (%)): 271.25 [M + H⁺]; HRMS-FAB (m/z [M + H⁺] Calcd; for C₁₅H₁₄N₂O₃: 270.29. Found: 270.25.



(E)-N'-(3,5-Dichlorobenzylidene)-2,6-dihydroxy-4-methylbenzohydrazide (**4e**). Yield: 75 %; m.p.: 130–131 °C; IR (KBr, cm⁻¹): 3082, 2954, 1687, 1595, 1519, 1466, 1385, 1332, 1276, 1217, 1152, 1097, 1020, 927, 860, 816, 769, 665; ¹H-NMR (400 MHz, CDCl₃, δ / ppm): 2.38 (3H, s, Me), 7.27–7.29 (2H, m, Ar-H), 7.41–7.42 (1H, m, Ar-H), 7.91–7.93 (1H, m, =CH), 8.12 (1H, s, NH), 9.37 (2H, s, Ar-OH); ¹³C-NMR (100 MHz, CDCl₃, δ / ppm): 20.4 (C-8), 104.4 (C-1), 112.1 (C-3/5), 127.9 (C-11/15), 129.9 (C-13), 134.6 (C-12/14), 136.3 (C-10), 138.7 (C-4), 142.3 (C-9), 159.5 (C-2/6), 173.4 (C-7); MS (EI, 70 eV, m/z (%)): 340.15 [M + H⁺]; HRMS-FAB (m/z [M + H⁺] Calcd for C₁₅H₁₂Cl₂N₂O₃: 339.17. Found: 339.15.

Fig. S-1. ¹H NMR (400 MHz, CDCl₃) spectrum of 1.Fig. S-2. ¹³C NMR (400 MHz, CDCl₃) spectrum of 1.

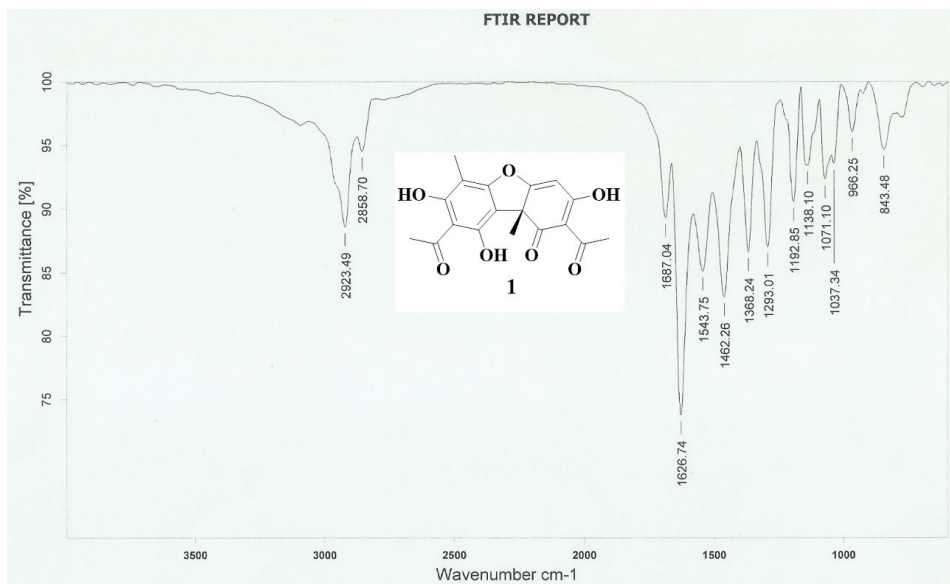


Fig. S-3. FT-IR (KBr) spectrum of **1**.

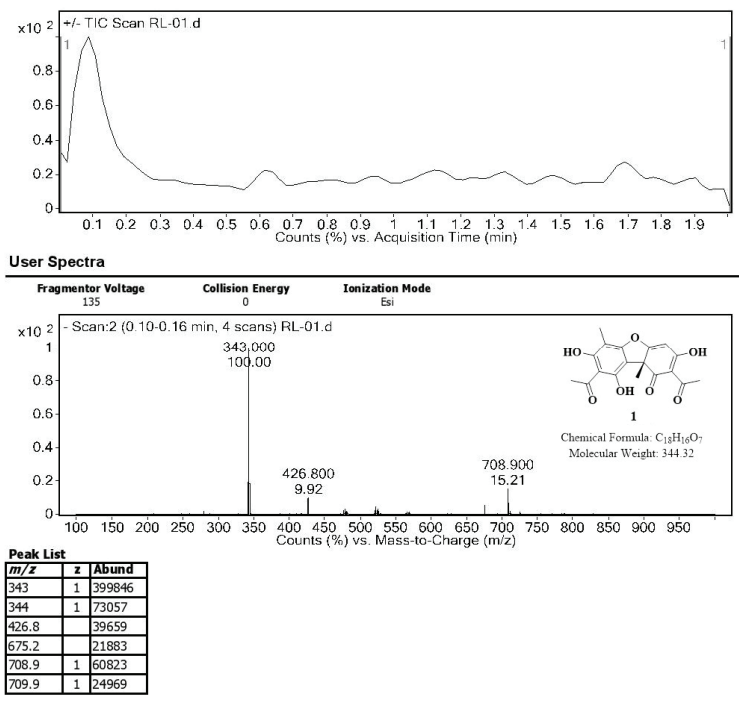
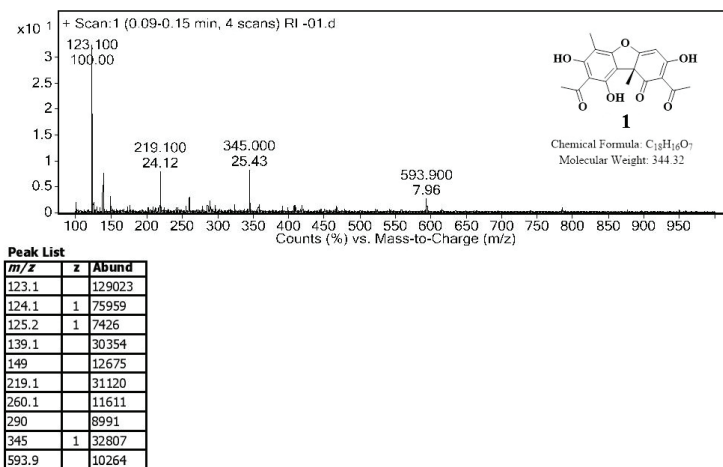
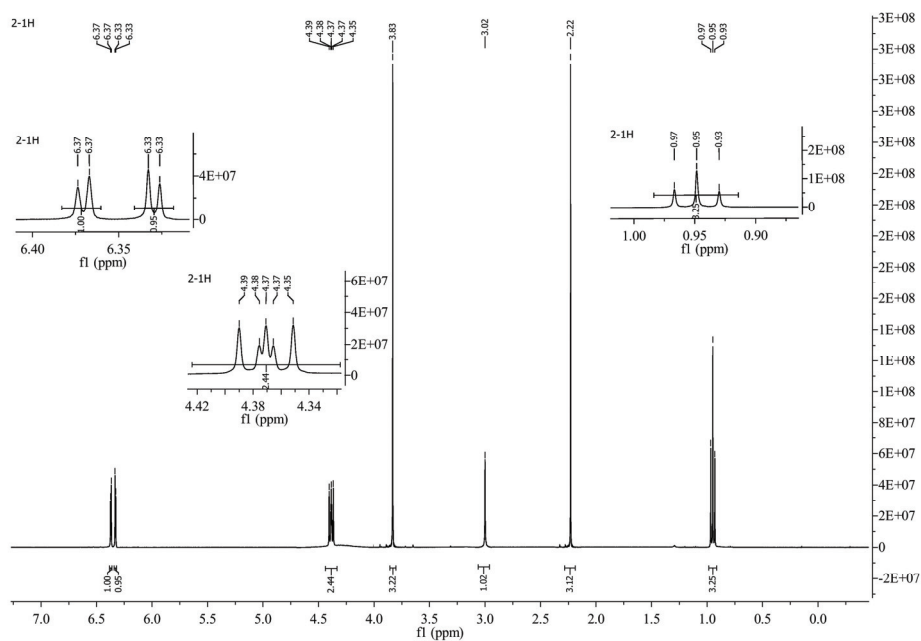
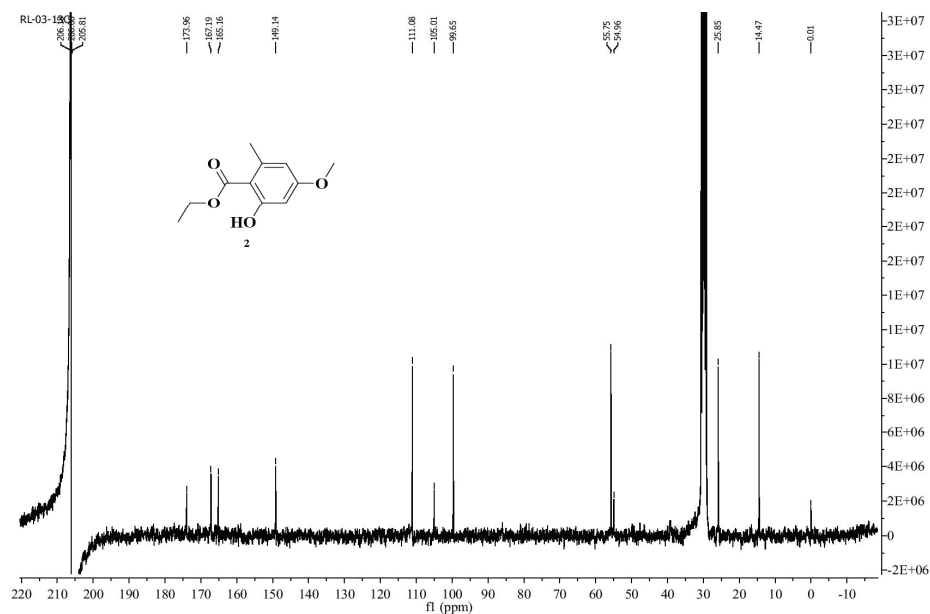
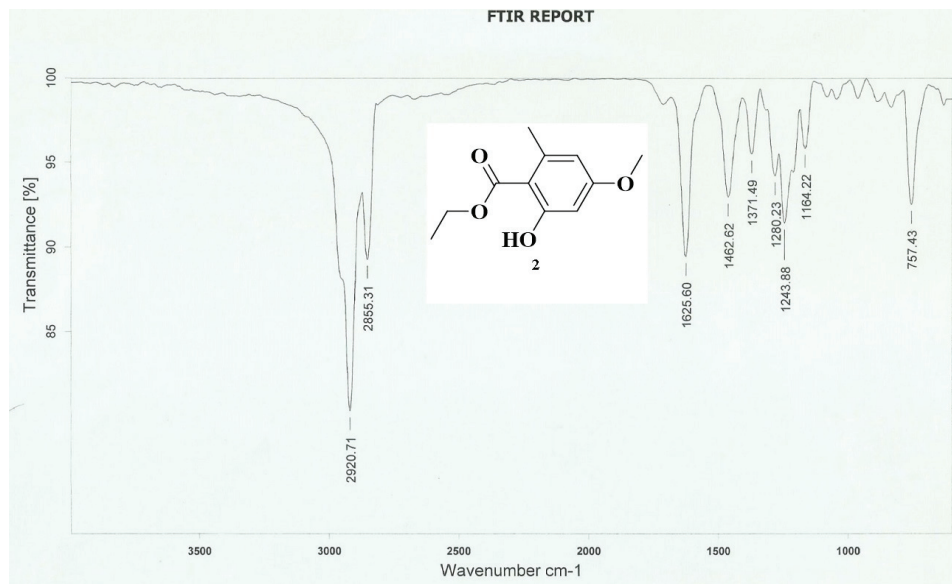


Fig. S-4. ESI-MS spectrum of **1**.

Fig. S-4a. ESI-MS spectrum of **1**.Fig. S-5. ¹H-NMR (400 MHz, DMSO-*d*₆) of **2**.

Fig. S-6. ^{13}C -NMR (100 MHz, acetone- d_6) of **2**.Fig. S-7. FT-IR (KBr) spectrum of **2**.

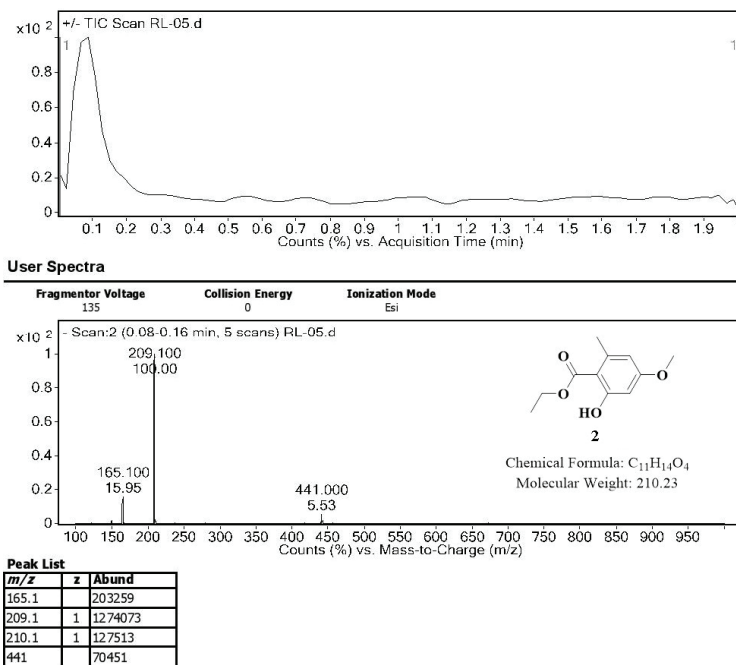


Fig. S-8. ESI-MS spectrum of 2.

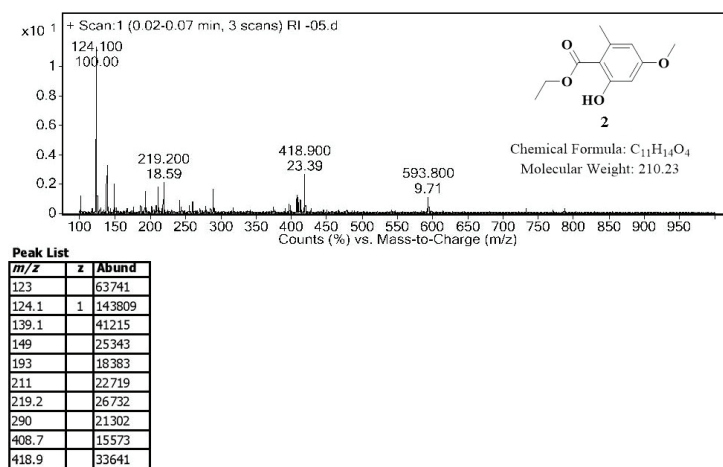


Fig. S-8a. ESI-MS spectrum of 2.

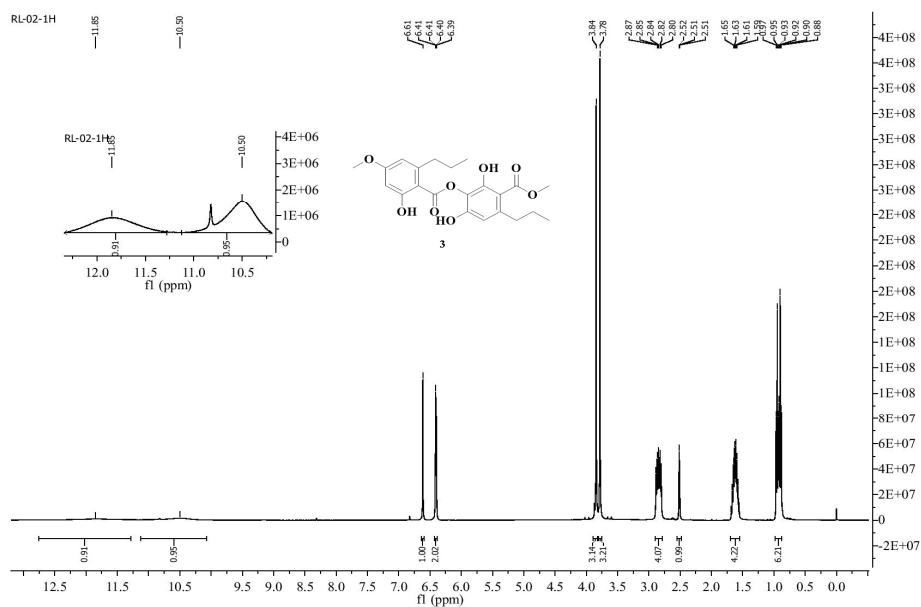


Fig. S-9. ¹H-NMR (400 MHz, DMSO-*d*₆) spectrum of **3**.

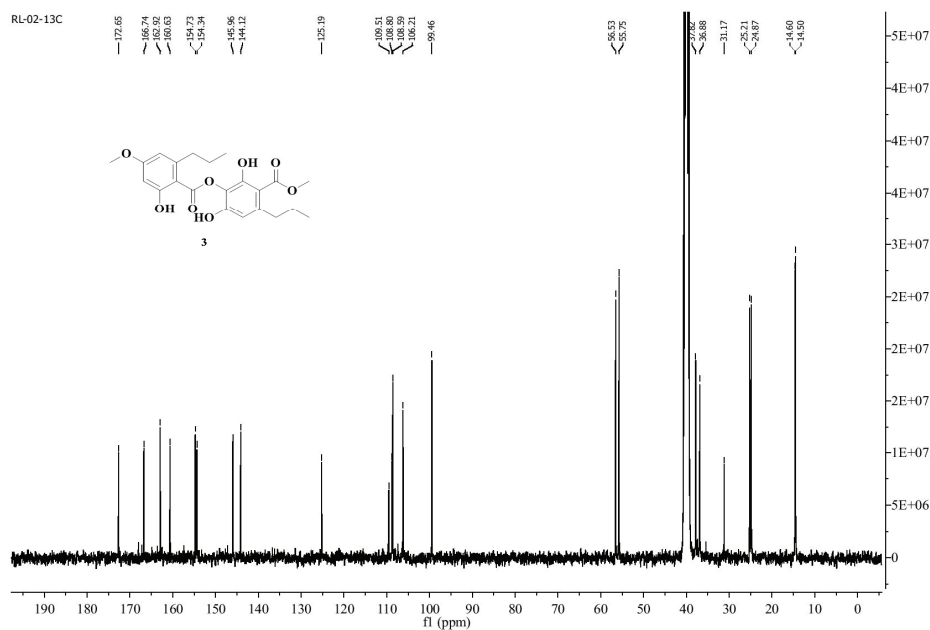
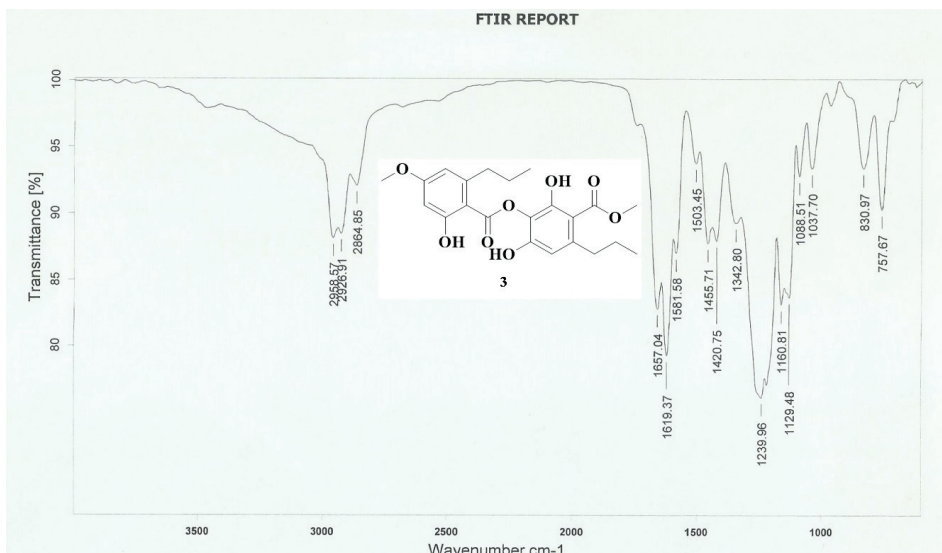
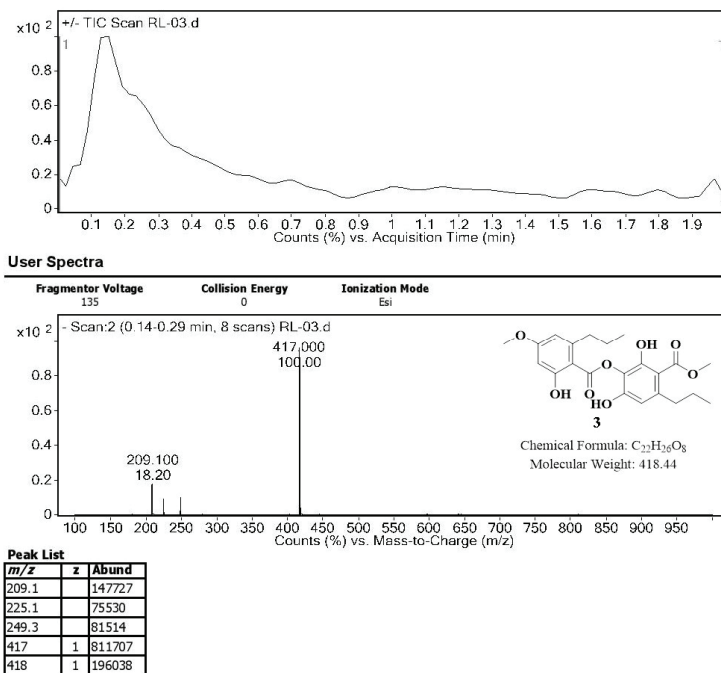


Fig. S-10. ¹³C-NMR (100 MHz, DMSO-*d*₆) spectrum of **3**.

Fig. S-11. FT-IR (KBr) spectrum of **3**.Fig. S-12. ESI-MS spectrum of **3**.

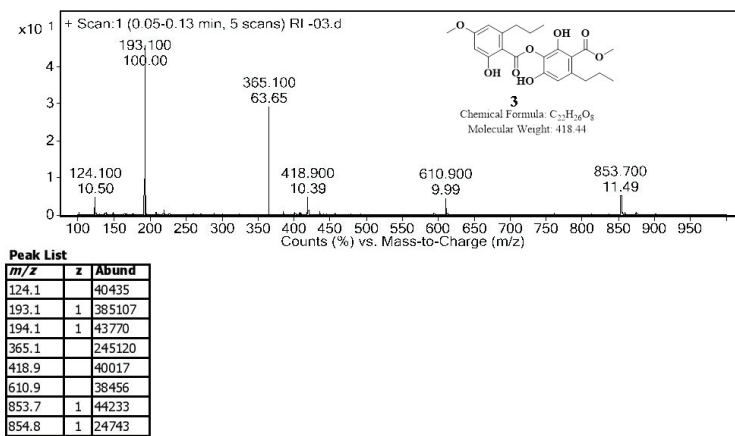


Fig. S-12a. ESI-MS spectrum of **3**.

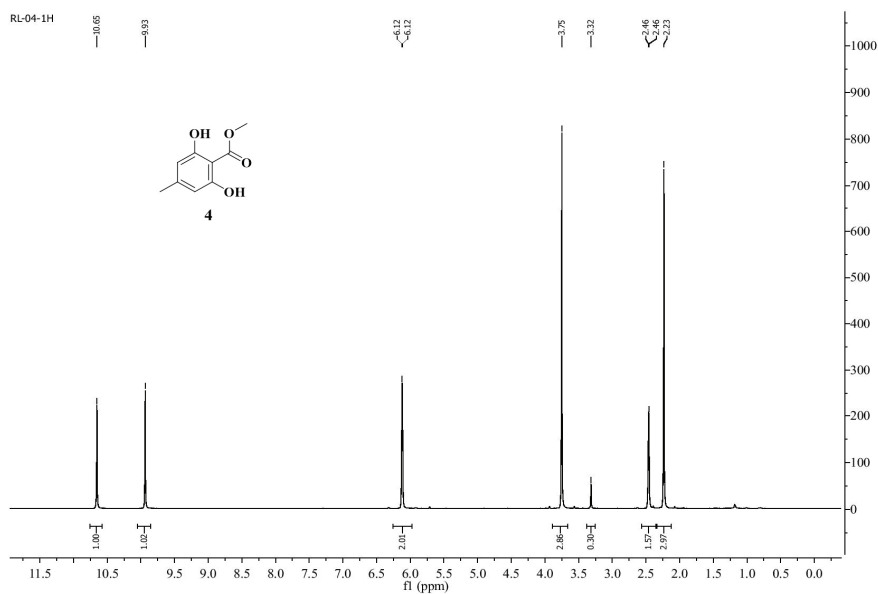
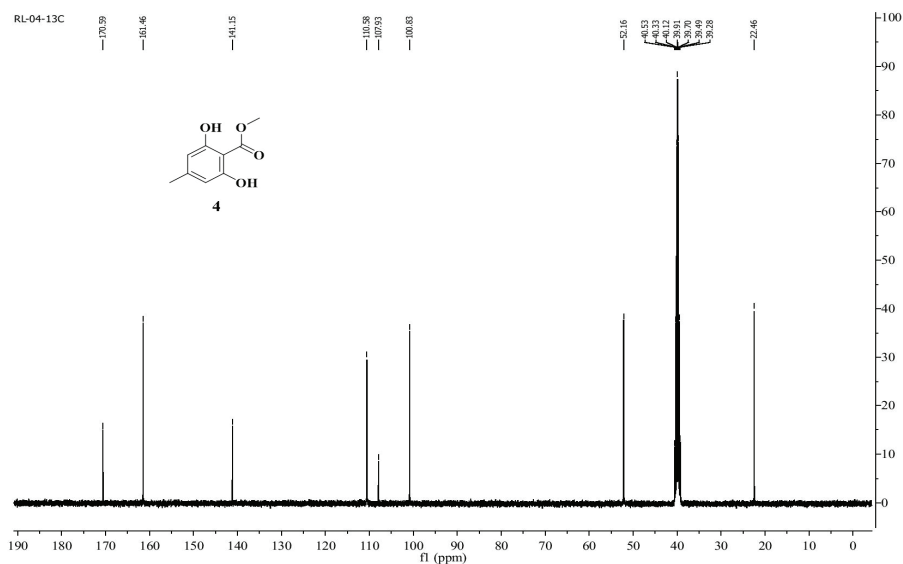
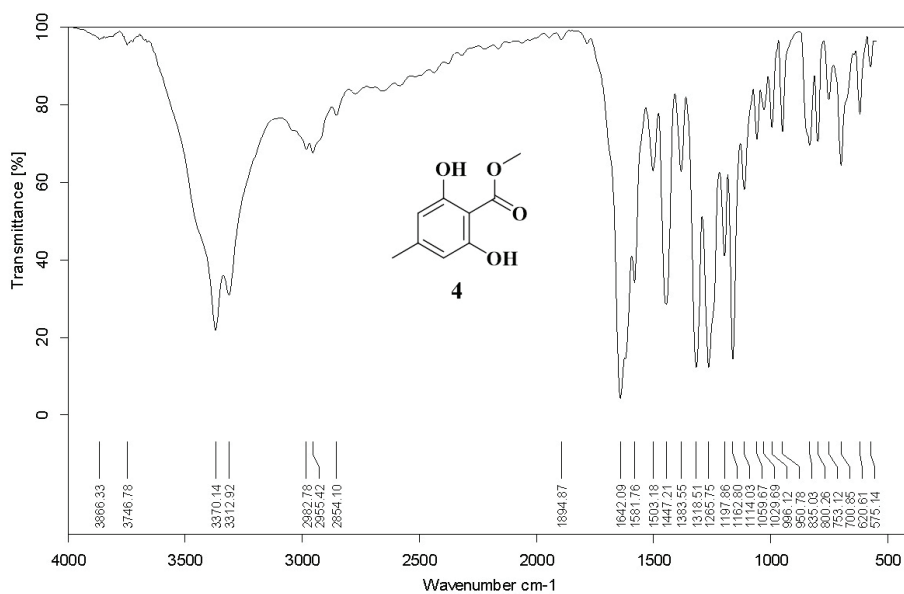


Fig. S-13. $^1\text{H-NMR}$ (400 MHz, $\text{DMSO-}d_6$) spectrum of **4**.

Fig. S-14. $^{13}\text{C-NMR}$ (100 MHz, $\text{DMSO-}d_6$) of **4**.Fig. S-15. FT-IR (KBr) spectrum of **4**.

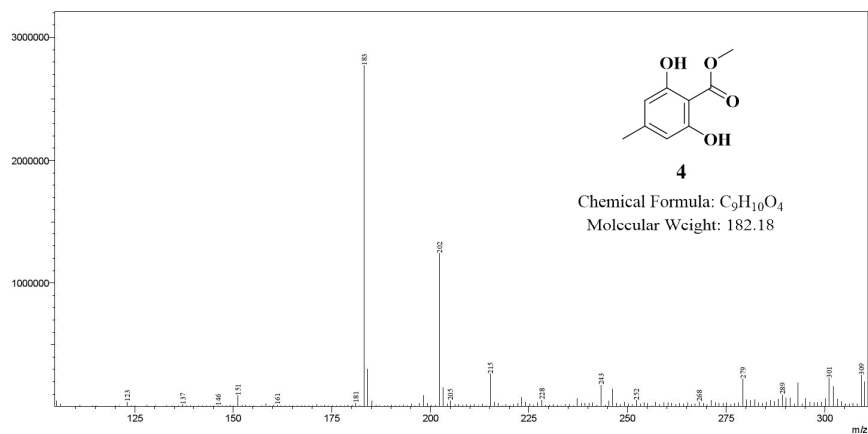


Fig. S-16. ESI-MS spectrum of **4**.

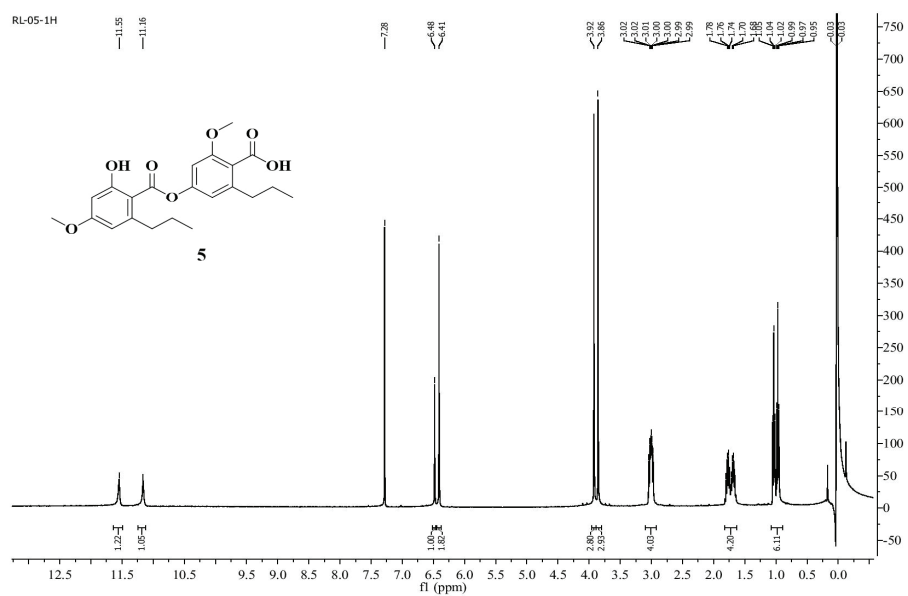


Fig. S-17. ¹H-NMR (400 MHz, CDCl₃) spectrum of **5**.

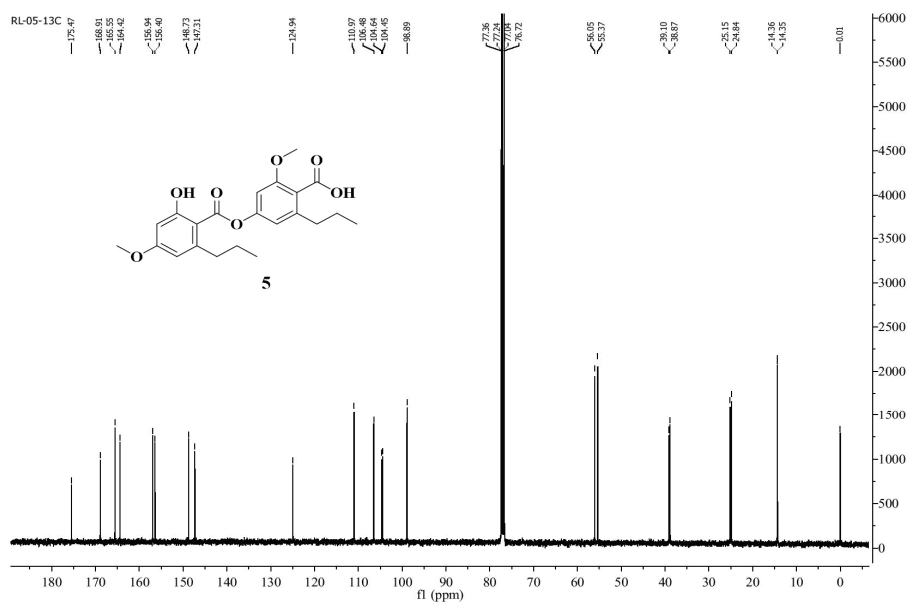
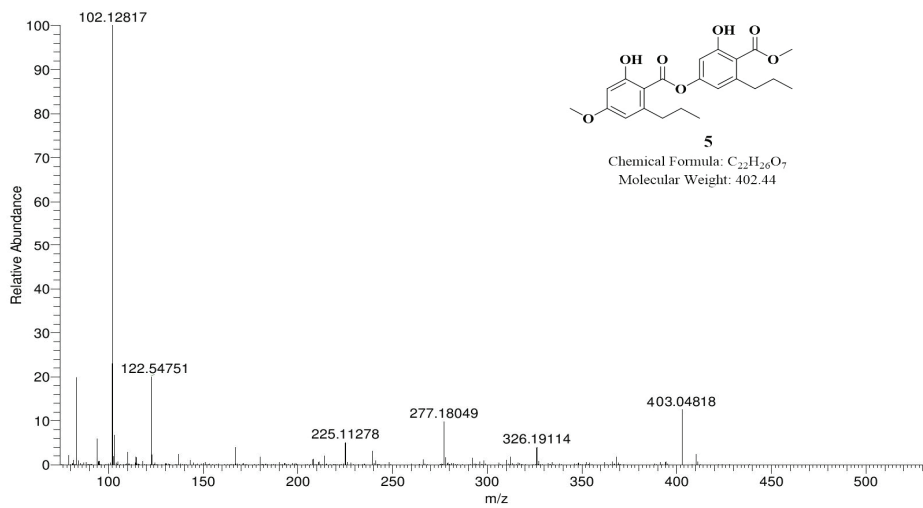
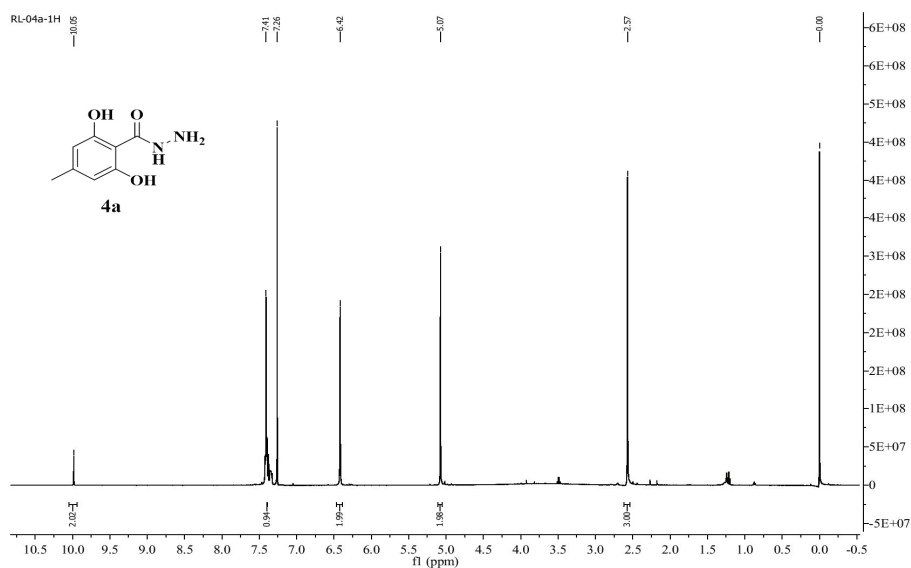
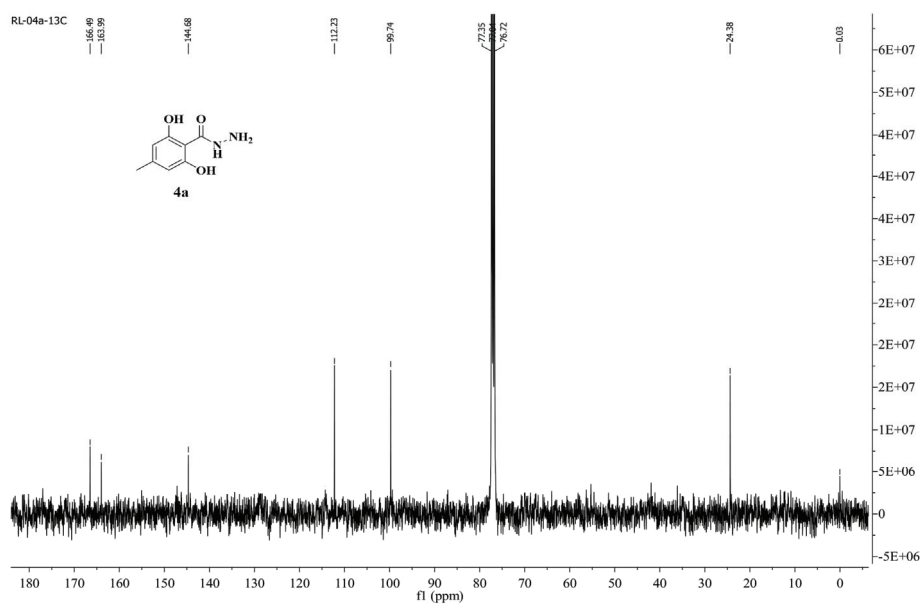
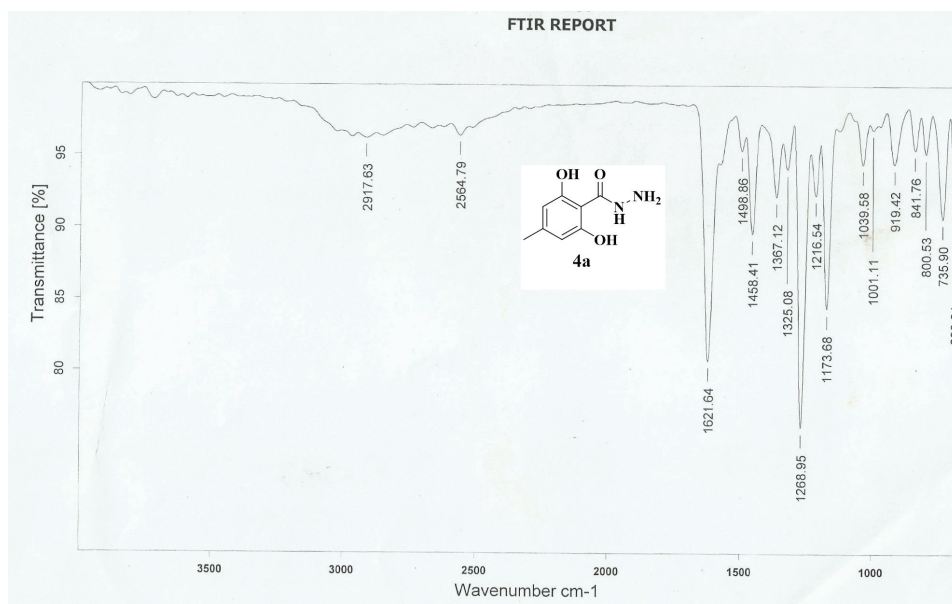
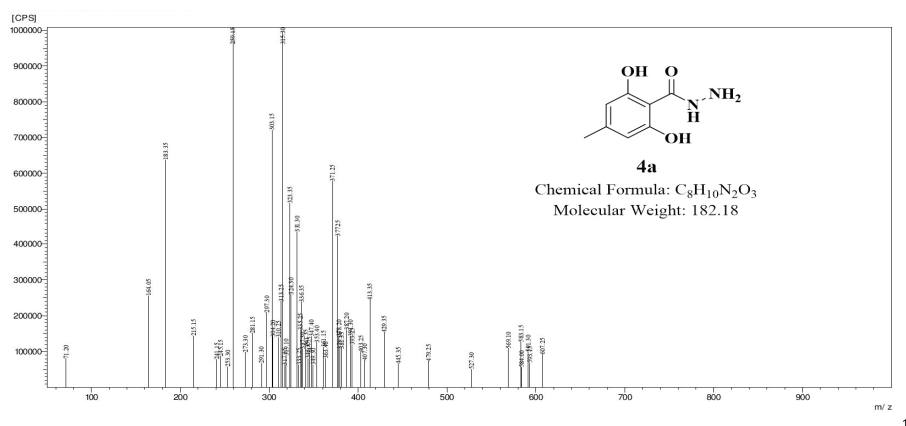
Fig. S-18. ^{13}C -NMR (100 MHz, CDCl_3) spectrum of 5.

Fig. S-19. ESI-MS spectrum of 5.

Fig. S-20. ¹H-NMR (400 MHz, CDCl₃) spectrum of 4a.Fig. S-21. ¹³C-NMR (100 MHz, CDCl₃) spectrum of 4a.

Fig. S-22. FT-IR (KBr) spectrum of **4a**.Fig. S-23. ESI-MS (positive mode) spectrum of **4a**.

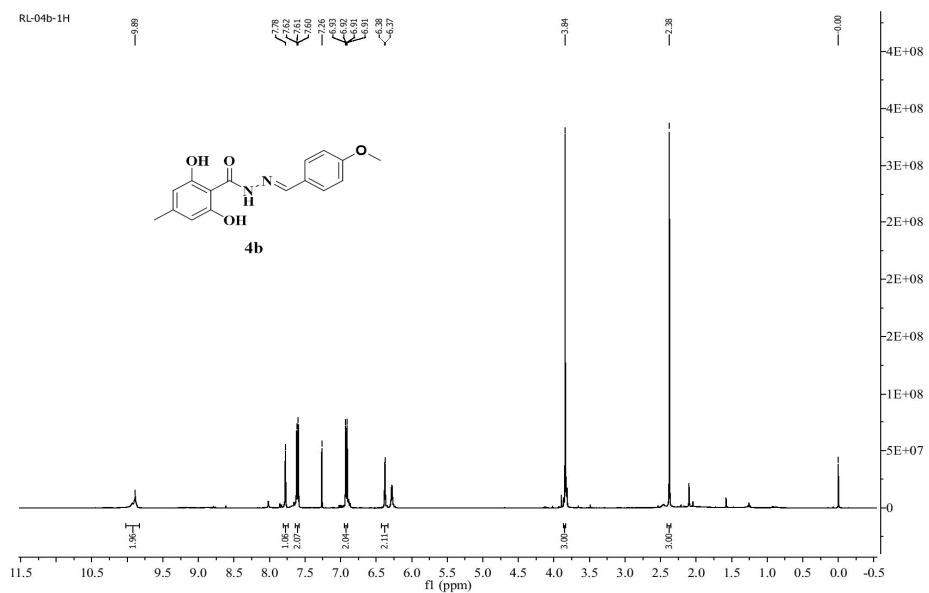


Fig. S-24. ¹H-NMR (400 MHz, CDCl₃) spectrum of **4b**.

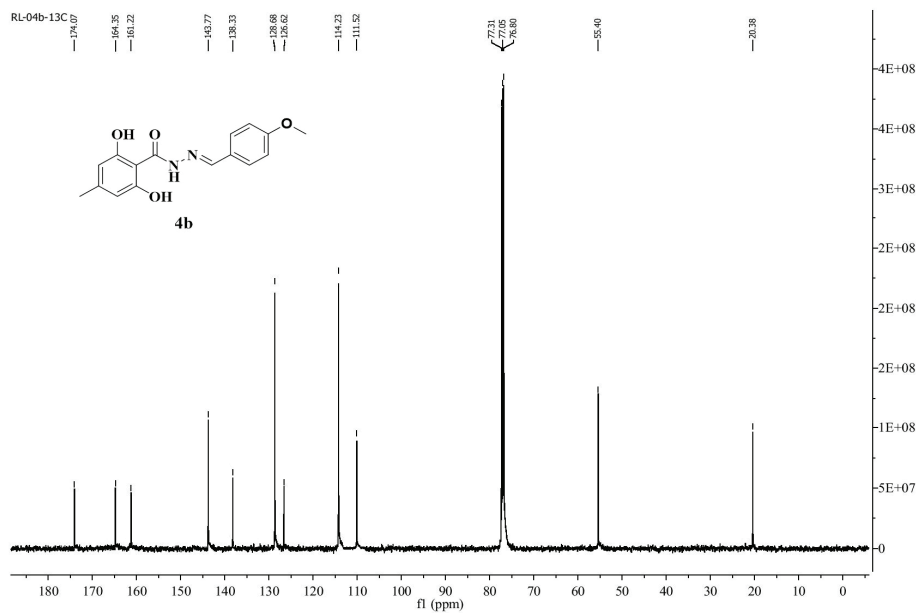
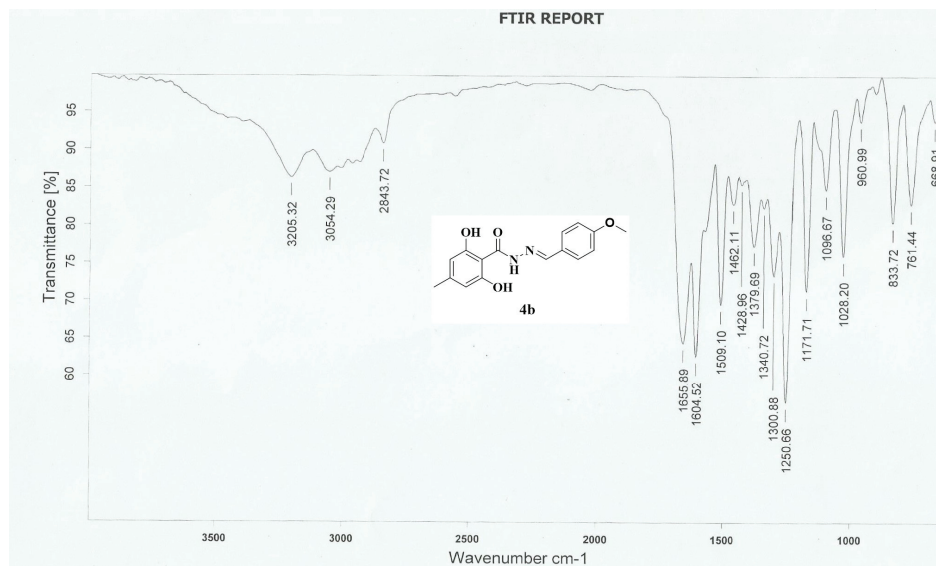
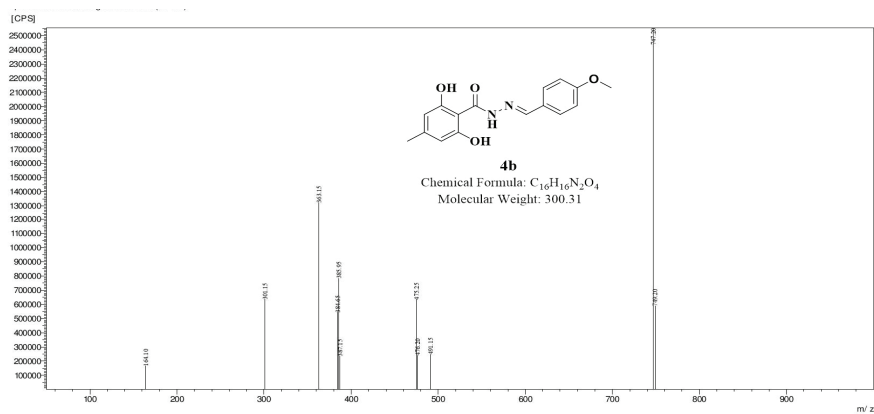


Fig. S-25. ¹³C-NMR (100 MHz, CDCl₃) spectrum of **4b**.

Fig. S-26. FT-IR (KBr) spectrum of **4b**.Fig. S-27. ESI-MS (positive mode) spectrum of **4b**.

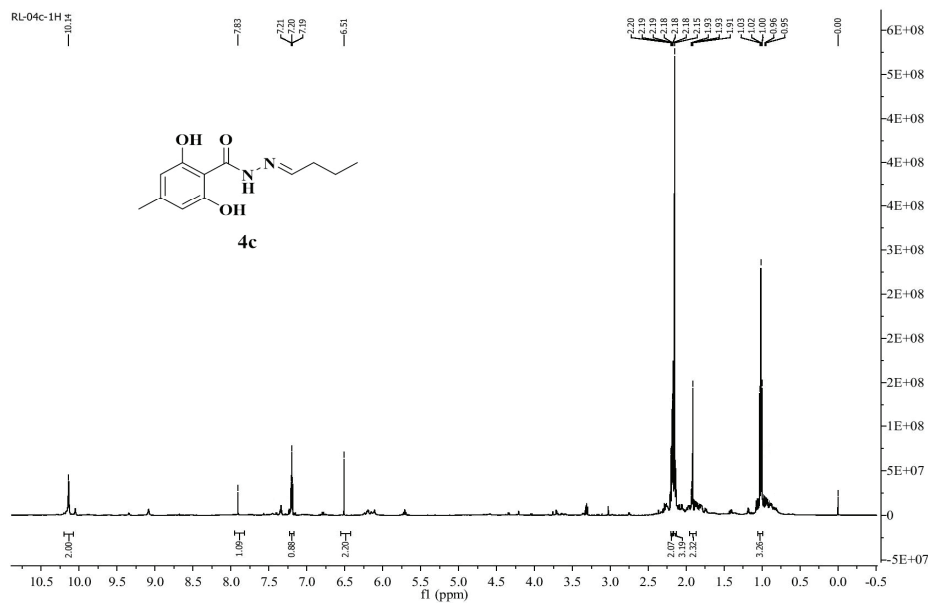


Fig. S-28. ¹H-NMR (400 MHz, CDCl₃) spectrum of **4c**.

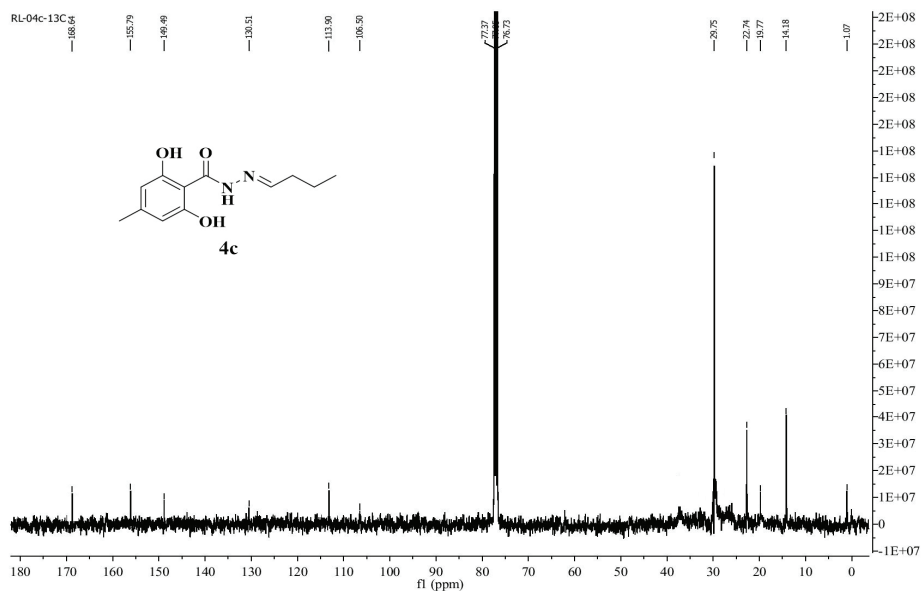
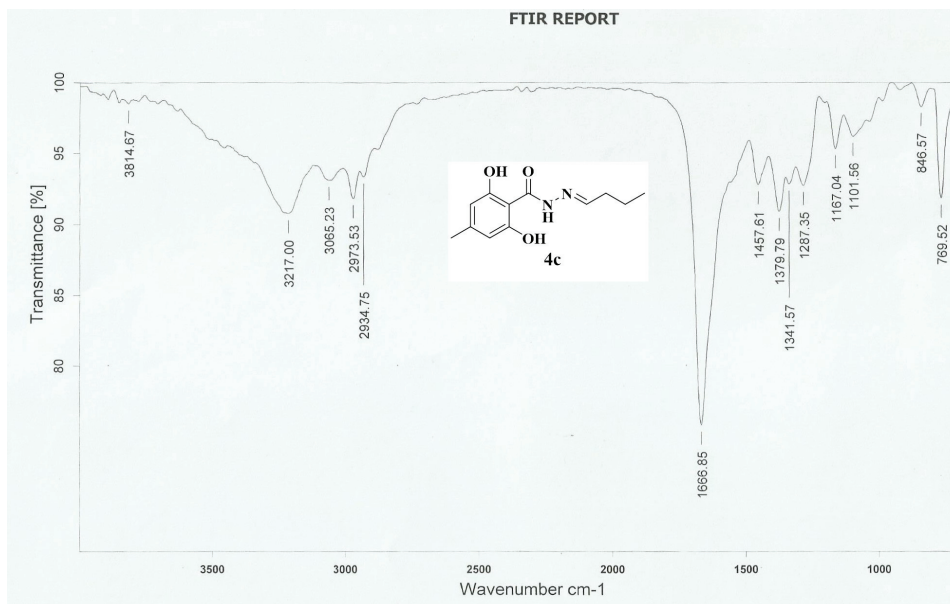
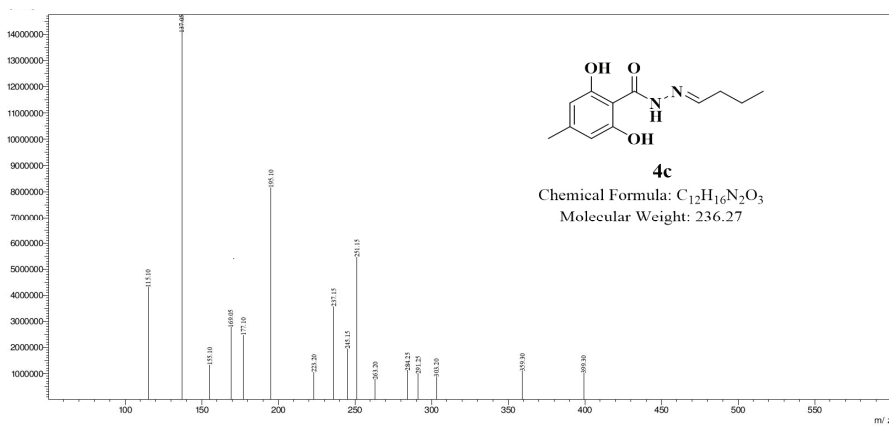


Fig. S-29. ¹³C-NMR (100 MHz, CDCl₃) spectrum of **4c**.

Fig. S-30. FT-IR (KBr) spectrum of **4c**.Fig. S-31. ESI-MS (positive mode) spectrum of **4c**.

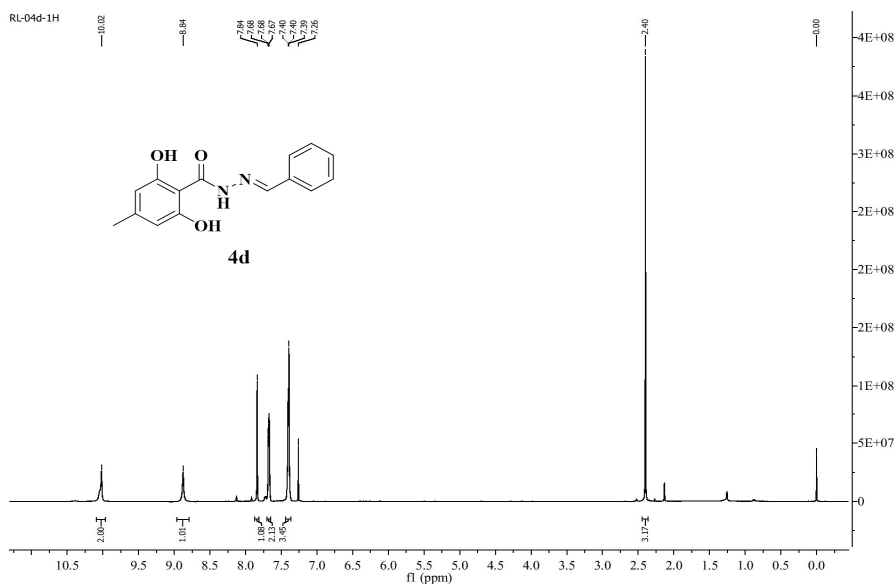


Fig. S-32. $^1\text{H-NMR}$ (400 MHz, CDCl_3) spectrum of **4d**.

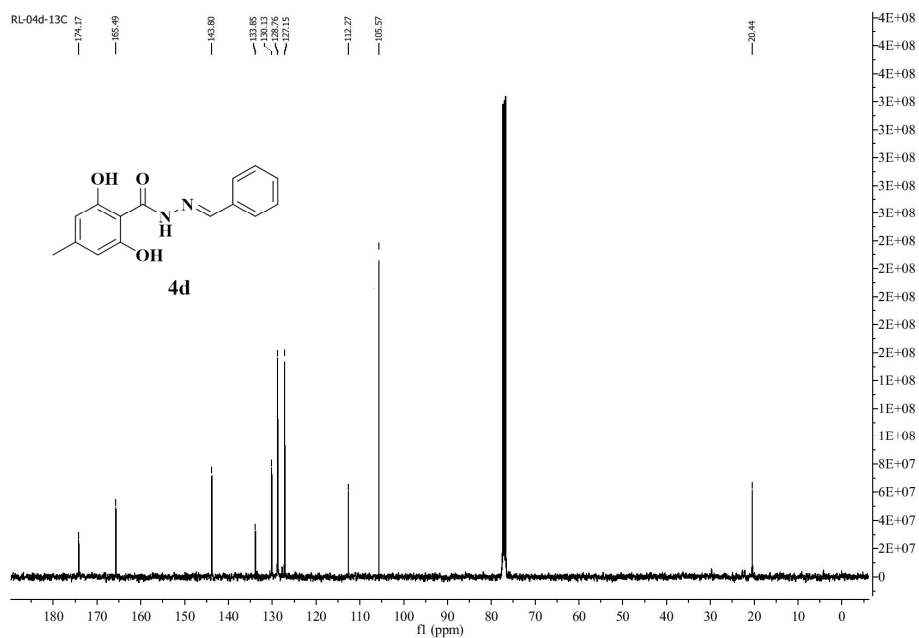
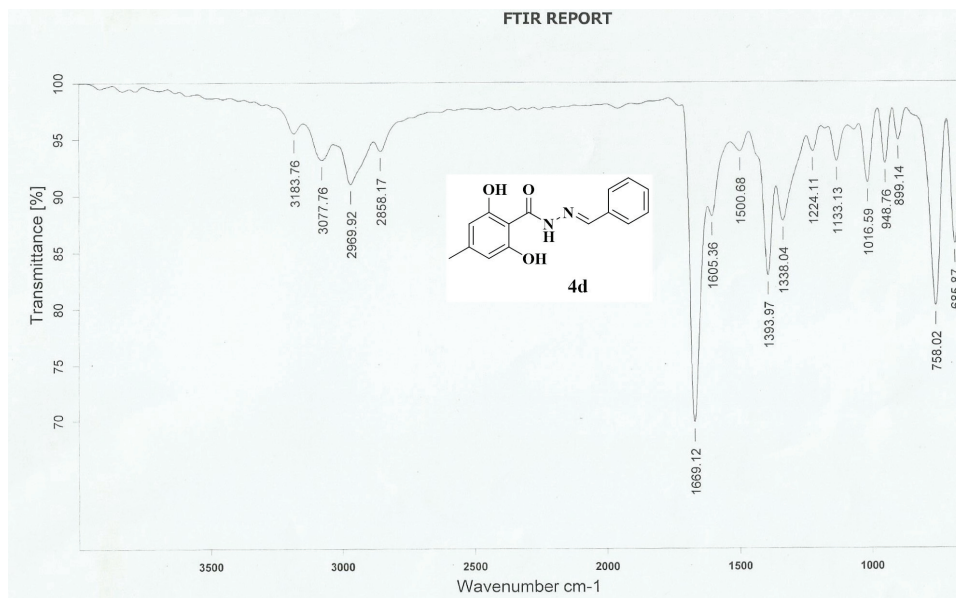
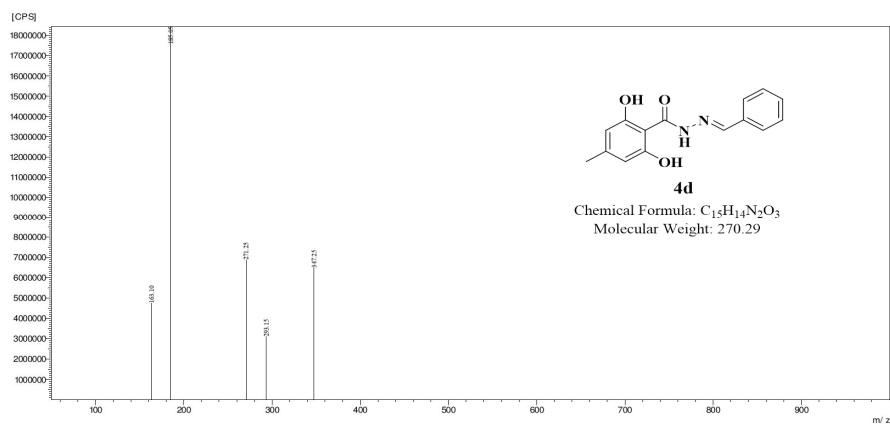


Fig. S-33. $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) spectrum of **4d**.

Fig. S-34. FT-IR (KBr) spectrum of **4d**.Fig. S-35. ESI-MS (positive mode) spectrum of **4d**.

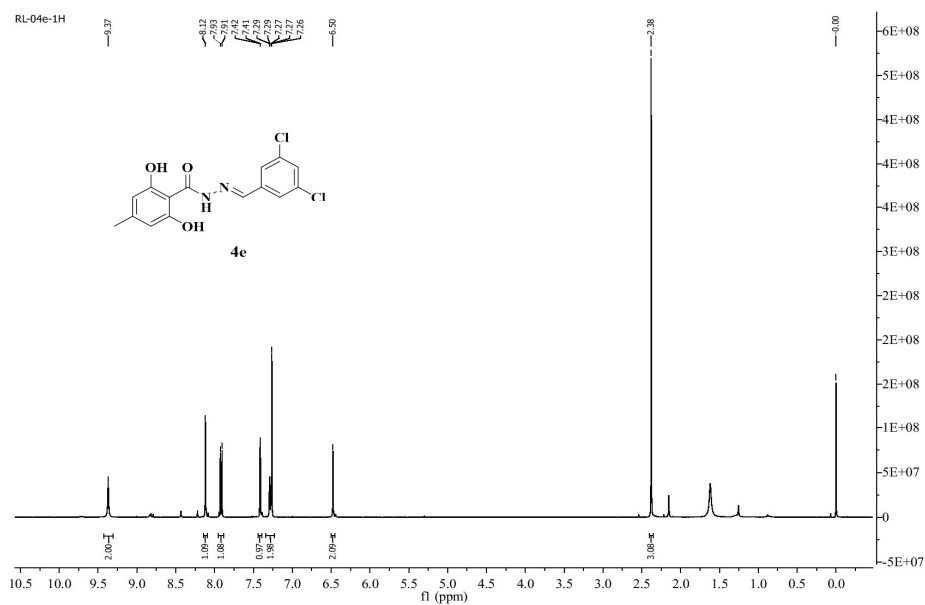


Fig. S-36. $^1\text{H-NMR}$ (400 MHz, CDCl_3) spectrum of **4e**.

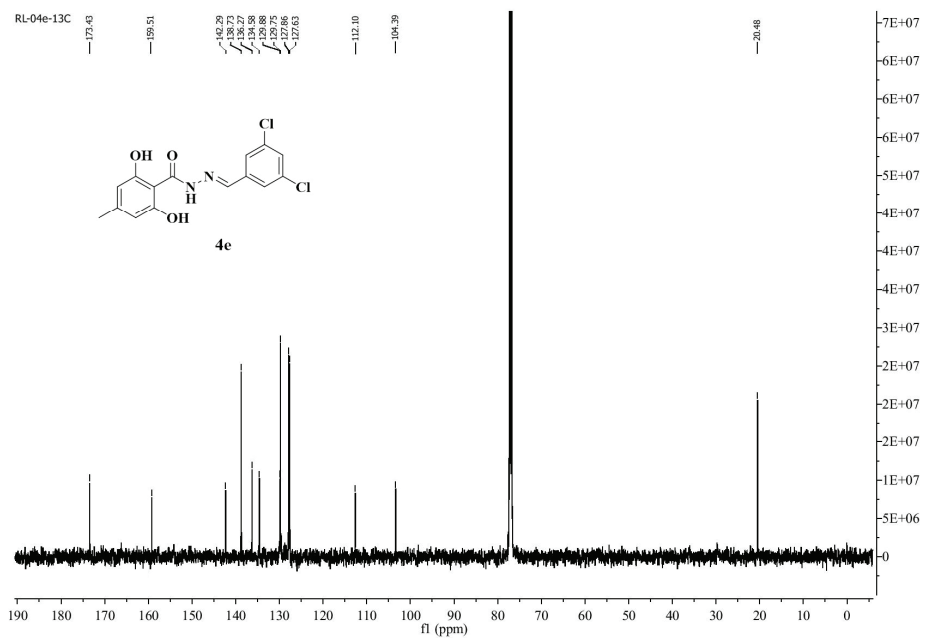
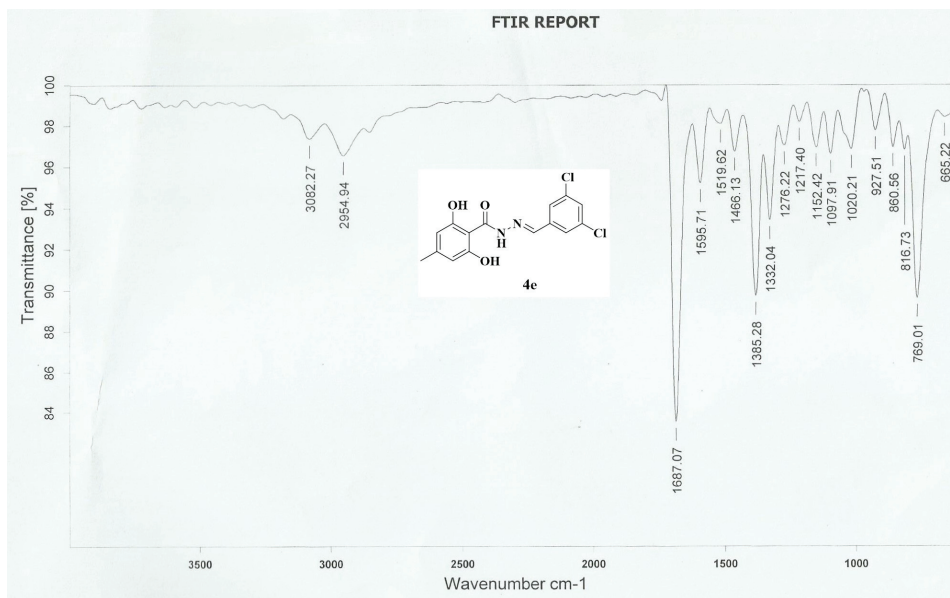
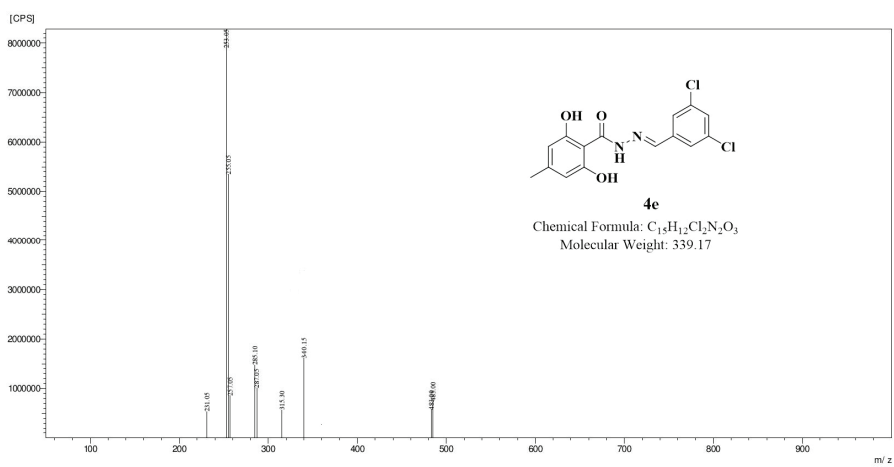


Fig. S-37. $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) spectrum of **4e**.

Fig. S-38. FT-IR (KBr) of **4e**.Fig. S-39. ESI-MS (positive mode) spectrum of **4e**.

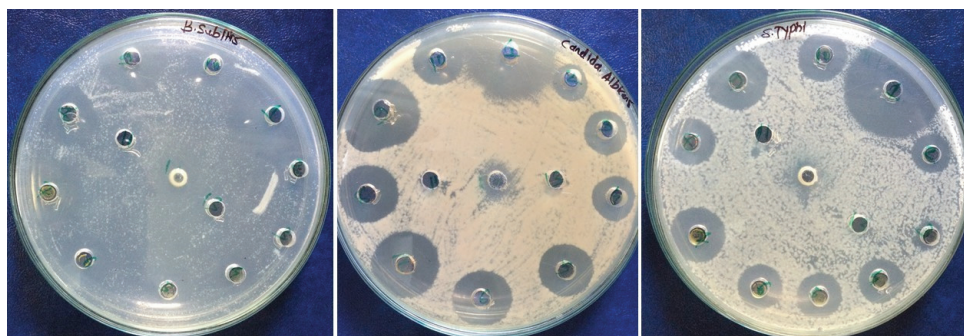


Fig. S-40. Zones of inhibition of 1–5 and 4a–e against bacterial and fungal stains.

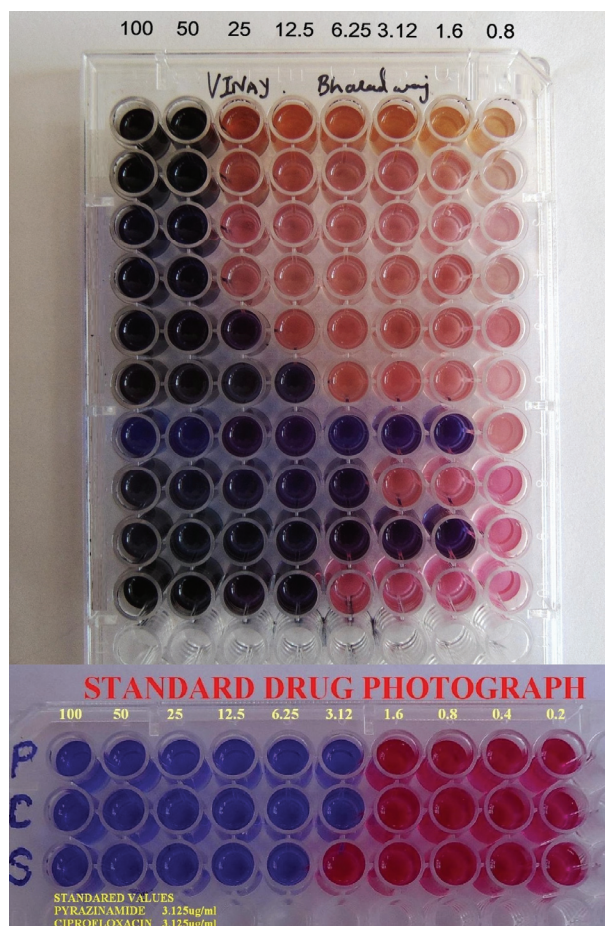


Fig. S-41. *In vitro* anti-tubercular activity of all the isolates and benzohydrazide derivatives of *Ramalina leiodea* against the *Microbacterium tuberculosis* H37Rv strain.

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3. K. R. Paidi, V. B. Tatipamula, M. K. Kolli, V. R. Pedakotla, *Int. J. Chem. Sci.* **15** (2017) 172 (<https://www.tsijournals.com/articles/benzohydrazide-incorporated-imidazo-12b-pyridazine-synthesis-characterization-and-in-vitro-antitubercular-activity.pdf>).